

Manual FASIMU

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Contents

1	Introduction	4
1.1	Layer-structure	4
1.2	The logical structure of a FASIMU session	5
1.3	General guidelines of FASIMU's user interface	6
2	Installation	8
2.1	Required unix tools	8
2.2	Further useful tools for FASIMU	9
2.3	Optimization solver	9
2.4	Install FASIMU	12
2.5	Visualization packages	12
3	FABASE-models	14
3.1	Getting a model from CellNetAnalyzer	14
3.2	Getting a model from SBML	14
3.3	Getting a model from a reaction scheme	15
3.4	Writing a FABASE model from the scratch	15
3.5	Refining the FABASE Model	16
3.6	FASIMU-compatible Models	19
3.7	Modify model files in a FASIMU session	19
4	The steps in a FASIMU session	21
4.1	Starting FASIMU	21
4.2	The control file <code>simulations</code>	21
4.3	Controlling the computation function	24
4.4	Perform simulations: <code>simulate</code>	28
4.5	The evaluation file: <code>evaluation.txt</code>	29
4.6	The comprehensive solution file: <code>allout.txt</code>	29
4.7	Prepare files for the visualization	30
5	Further functions in FASIMU	31
5.1	Prune to <i>ad functionem</i> networks: <code>prune-network</code>	31
5.2	Flux-variability analysis	31
5.3	Flux control analysis	32
5.4	Check the essentiality for a given set of simulations: <code>check-essentiality</code>	32
5.5	Functions creating common <code>simulations</code> files	33
5.6	Functions creating common exchange files referenced in a <code>simulations</code> file	34
5.7	Network printing functions	34

6	Functions for sets of modes: <code>modeset</code>	36
6.1	Creating a <code>modeset</code> from <code>allout.txt</code> and vice versa	36
6.2	Which mode fits a given profile best? <code>modeset-score</code>	37
6.2.1	Zeros in the mode M_k <code>sigscore</code> (-S) vs. <code>fullscore</code>	37
6.2.2	Scaling factor: fixed (-f), dependent on m_z (-L), or computed by linear regression . .	37
6.2.3	Relative (-R) vs. absolute	38
6.2.4	Cumulative (-C) vs. fitting	38
6.2.5	Setting the center of the score distribution manually (-c)	38
6.2.6	Setting the steepness of the distribution function (-s)	38
6.2.7	Should the score variance depend on the flux value (varsingle -v)?	39
6.2.8	Mathematical description of the score distribution (-t)	39
7	Using FABASE without FASIMU	43
7.1	Computation functions	43
7.2	Further functions in FABASE	45
7.3	Modify model files in a FABASE session	48
8	Customization of the external solvers	50
9	Final remarks	52
9.1	Publication	52
9.2	Getting more help	52
9.3	License	53

Chapter 1

Introduction

FASIMU is an extremely flexible computation environment for different sorts of flux-balance analysis. The flexibility is traded off with simplicity of use. However, people who are acquainted with command-line interfaces will find it easy to use. On the pro side is that practically any algorithmic parameter can be adjusted and it is therefore especially suited for the scientific workflow: difficult problems can be tackled interactively and session protocols can be used to implement new flux-balance methods.

FASIMU is very open in two aspects: The source code is open and written in a high-level language which makes it easy to adapt and implement new functions. The other aspect is that intermediate results are stored in human-readable files which can easily be analyzed once the internal structure of the workflow is understood.

The interactive language used for FASIMU is one that most LINUX and MacOS users along with many other UNIX and Windows users are familiar with: the Bourne again shell (bash) [1]. Thus, anything working in bash can easily be combined with FASIMU. The typical FASIMU session is in every respect identical to the interactive work in bash. In other words, FASIMU consists of available bash-functions. This concept is more flexible than the interactive languages of Mathematica or MATLAB as the integration with other programs is much more straightforward. For users of LINUX the learning curve is very steep for the syntax of the available commands. Writing algorithmic ideas based on FASIMU is technically writing additional bash functions. Programs in other languages can this way easily be combined with FASIMU; the only prerequisite is an executable program and a defined file exchange.

In flux balance analysis by far the most computational expensive part is the work of the optimizer. Thus, it is not much of an advantage to write the control of the algorithm in a low-level language. This is the reason that the implementation of FASIMU is consequently divided into two parts: the computationally difficult part is handled with an external solver (currently CPLEX, LINDO, lp_solve, or GLPK), the computationally easy but semantically complex part is written in a high-level language which is easiest to understand and modify.

1.1 Layer-structure

FASIMU consists of a two-layer structure: the outer layer is FASIMU itself and the lower layer is called FABASE. Where FABASE is focused on a single flux optimization, FASIMU supplies the iteration of flux optimizations with FABASE. FABASE can be used on its own when only single flux computations are required, FASIMU depends on FABASE. It is worth to keep in mind which function belongs to which level. The crucial item of the interface between the two layers is the shell variable “`$optimization_call`”. It contains a FABASE function with full parameters which is called for any of the FASIMU functions which iterate flux calculations.

There is also another respect in which FASIMU sets a layer structure: on the metabolic model itself.

When initially called, FASIMU modifies the model by setting a new boundary around the system and the addition of virtual exchange processes for any metabolite of the system across this boundary. The result is another FABASE-compatible model but with a few special properties. Usually, this works unattended in the background but it appears in the computed flux solutions. The duplicated metabolites can be recognized at the “_ex” ending. The virtual exchange processes hold a “_tr” suffix.

1.2 The logical structure of a FASIMU session

A FASIMU session should be started in a single empty directory. This directory will later contain the various input, output, and intermediate files of this particular session. For the analysis and proper reference it is advisable not to mix files from different sessions or edit the files in this place. FASIMU will probably work anyway but it is more difficult to assess if it is really doing what the scientist intended to do.

- The first step is to copy all files related to the metabolic network into that directory. It is advisable to store the information to one model version in a separate directory. The metabolic model must be supplied in FA-Format, this is a proprietary extended version of CellNetAnalyzer-Format. There are transformation tools available for SBML and a simple Reaction-Scheme-Format. See chapter 3.
- The second step is the call of “**source fasimu**” which has several purposes. It transforms to model into a form compatible with FASIMU and makes the FASIMU functions available. Then it starts FABASE which supplies the FASIMU functions and writes some intermediate files which accelerate the preparation time for a single flux optimization call: the main parts of the input into the optimizer program is the prepared (in LPF-Files). See section 4.1.
- The third step (which also might be interchanged with the second) is the writing of the iteration scheme, a file with the fixed name “**simulations.txt**”. One line defines a single flux computation. See section 4.2.
- The fourth step is setting the variable “**\$optimization_call**” which defines the algorithm of the flux computation. See section 4.3.
- The fifth step is the start of the simulations with “**simulate**”. This function proceeds with all simulations, there are also functions which select a simulation etc. See section 4.4.
- The sixth step is to review the file “**evaluation.txt**” whether the reported results have been satisfactory. There is one line for each simulation holding a “+” if it obtained the desired result, “-” otherwise. See section 4.5.
- The seventh step is to view the flux distributions stored for all solutions in a file “**allout.txt**”. Single flux distributions are selected with “**allout-select <name of the simulation>**”. The I/O-fluxes are at the beginning and should be reviewed first whether the system boundary fluxes are as desired. See section 4.6.
- The eighth step is advisable if the flux distribution seems reasonable but too complex to check in text form: the visualization in BiNA [16], Cytoscape [14], or CellNetAnalyzer [15]. Basically there is the function “**allout2bina**” which creates directories of input files for the flux analysis plugin of BiNA, or the function **allout2valfiles**. See section 4.7.

All these steps are described in greater detail below.

1.3 General guidelines of FASIMU's user interface

Fixed file names

The model under investigation be FASIMU resides in **fixed file names**. This has the immediate consequence that in one directory there can only be one model at a time.

Shell variables

Some information related to the present model are stored in local variable names. It is therefore not possible to change the directory to another FASIMU model and continue there. It is necessary to call `source fasimu` again in this new directory. This call regenerates the contents of the variables `$rea`, `$met`.

Some variables are set to default values by `source fasimu` if they are not defined but the are not overwritten if they are already defined: `$optimization_call`, `$default_compartment`, `$solver`, `$names_in_allout`.

An important variable is called `$fabase_zeroflux_threshold`. Flux rates with a smaller absolute value than this number are considered to be zero.

File contents

All input files are whitespace-separated text files except for the file `simulations` where a two level separation is necessary: the primary separator is the tab and the secondary separator are space characters. FASIMU's structured main output files (`allout.txt` and `evaluation.txt`) are tab-separated. FABASE output files are whitespace separated.

File extensions

All files to be supplied by the user are text files with a specific name such as `reactions` without an extension. Some of these files are modified during the course of computation, their original content is kept in `<file-name>.original`. All files produced by FASIMU being of interest to the user have the extension `txt` or `out`. Several files are produced during the workflow. They are recognized by their extension:

Extension	Function
none	input file
original	backup of input files
txt	produced by FASIMU being of interest to the user
fgf	internally used by FASIMU
lpf	parts of the problem formulation
lp	CPLEX-LP format
ltx	LINDO-LP format
par	parameter files of LINDO and <code>lp_solve</code>
in	Input to external programs
out	Output of external programs and extracts thereof

Function names

Function names carry dashes and not underline characters (except if it contains the identifier `lp_solve` which is dictated by the name of the solver)

File parameter

Functions do not accept file parameters, instead the files used have fixed names. If several files for the same purpose are used throughout a session, the session script must rename them. Few exceptions are `compute-moma` and `compute-room` which take a reference solution as a parameter.

Switch function upon file existence

Many functions are switched on if the respective file exists: e.g. fluxes are fixed if the file `setfluxes` just exists. Some files are used only for certain algorithms, such as `fluxmin-weights` is used only for flux minimization. Other files are only used if explicitly switched on such as `expressions`.

Switch functions upon file existence	
used for all optimizations	<code>fluxbounds</code> <code>setfluxes</code> <code>fluxfix</code>
used for flux minimization	<code>fluxmin-excluded</code> <code>fluxmin-weights</code>
used for thermodynamic constraint	<code>TR-excluded</code> <code>concentration-class-ranges</code> <code>concentration-ranges-classes</code>
must be switched explicitly	<code>expressions</code> <code>enzymes</code>
only in FABASE session	<code>fluxconstraints</code> <code>targetfluxes</code>

File overwrite

As a consequence of fixed file names, the invocation of FASIMU with `source fasimu` changes the input files for FABASE: `reactions`, `metabolites`, `equilibriums`, `targetfluxes`. The original files are kept in respective files with the extension `.original`. This concept has historical reasons (FABASE was first, FASIMU has been set on top of it). The parameter files (`cplex-head.in` and files with the extension `par`) are only created by `source fabase` / `fasimu` if not present. Present files will only be overwritten if explicitly requested by the `restore-default-parameters` function. Also, the function `set-timeout` modifies these parameter files.

Effects of source fasimu	
<code>reactions</code>	modified, backup in <code>reactions.original</code>
<code>metabolites</code>	modified, backup in <code>metabolites.original</code>
<code>equilibriums</code>	modified, backup in <code>equilibriums.original</code>
<code>fluxmin-excluded</code>	modified, backup in <code>fluxmin-excluded.original</code>
<code>TR-excluded</code>	modified, backup in <code>TR-excluded.original</code>
<code>concentration-ranges-classes</code>	modified, backup in <code>concentration-ranges-classes.original</code>

During the FASIMU functions `simulate` and `simulate-single`, the files `targetfluxes` and `fluxconstraints` are overwritten and should not be modified by the user. In a FASIMU session, `setfluxes` and `fluxbounds` should be used instead.

Choice FASIMU vs. FABASE session

By starting the session with

```
| source fasimu
```

you have all features of FASIMU but you can also start the so-called FABASE session with

```
| source fabase
```

which give more manual control options. See chapter 7 (Using FABASE without FASIMU).

Chapter 2

Installation

FASIMU requires some basic UNIX tools and an optimization solver. Also, a visualization package increases its usefulness.

2.1 Required unix tools

Bash

On computer systems based on LINUX and MacOS, bash is installed as part of the operating system. On UNIX systems, where another default shell is used and bash is not yet available on the installation medium, it can be compiled from the freely available source code, see <http://www.gnu.org/software/bash>. The version of bash should be at least 4.0 for the function `prune-network` to work, the rest of FASIMU also works for older versions.

Gawk

On LINUX systems GNU AWK (gawk) is installed as part of the operating system but it is preinstalled on many other UNIX and MacOS systems. If it is not preinstalled it can usually be installed from the installation source. If that is not possible it can be compiled from the freely available source code at <http://www.gnu.org/software/gawk>. The version of gawk should be at least 3.0.

GNU AWK is an implementation of a more generally defined language AWK and has some extensions. FASIMU uses some of the extensions, thus, other implementations of AWK might not work for FASIMU. Therefore it is not a good practice to link or rename gawk to awk. FASIMU calls gawk explicitly. Call

```
| gawk -W version
```

from the command line to check if that works.

The actual position of the gawk executable in the file system may vary in certain UNIX distributions. FASIMU assumes `/usr/bin` as the position of gawk but it may also be `/bin`. If that is the case, the script `sbml2fa` and `reaction2fa` may result in an Error message “`/usr/bin/gawk not found`”. The easiest way then is to call it with:

```
| gawk -f /usr/local/bin/sbml2fa <SBML-file>
```

if the install directory of FASIMU is `/usr/local/bin`.

Other UNIX commands

FASIMU uses other basic UNIX commands: `cat`, `grep`, `egrep`, `fgrep`, `ls`, `echo`, `mv`, `cp`, `head`, `tail`, `wc`, `diff`. In very rare circumstances they are not preinstalled on certain systems. In that case, FASIMU reports an error “command not found” for this command, and you should find a way to install this command.

Unix tools on Windows

On Microsoft Windows systems `bash` and `gawk` are usually not installed. They can be conveniently installed by installing Cygwin. The recent version of Cygwin supports, Windows NT from version 4.0, Windows 2000, Windows XP, Windows Server 2003, Windows Server 2008, Windows Vista, Windows 7. Install a cygwin system by the calling the URL <http://www.cygwin.com/setup.exe> in a browser window and execute the setup program. In the process of installing you are asked to enter a directory to install cygwin in and a mirror to download software. Finally, you get a window to select the software to install. You can check whether `gawk` and `bash` are on the list of the software to install but that should normally be the case. You should also install `glpk` (in the Math directory), simply enter `glpk` in the search box and click to select for install.

On Windows 98, Windows ME, Windows CE an alternative version of cygwin can be installed, use the URL <http://www.cygwin.com/setup-legacy.exe> to install.

After the installation a program `Cygwin bash shell` is added to the start menu. Start that and you are in `bash` shell and can proceed to install FASIMU.

FASIMU on other architectures

There is no restriction for computer architectures as long as long the basic UNIX tools and a solver can be installed. However, this has not been tested so far.

2.2 Further useful tools for FASIMU

The UNIX tools `wget`, `unzip` (GNU zip), `less` are normally also installed on any system where also `bash` is installed. They are useful for the command-line work and are occasionally used in the examples in the manual and the tutorial.

The http-program `wget` is used to download FASIMU files from the bionformatics.org server. If you are protected by a firewall program which connects through the internet with a proxy, you have to set the variable `http_proxy`.

2.3 Optimization solver

FASIMU requires the installation of an optimization solver. It was originally developed with the commercial solver ILOG CPLEX, but also the free solvers `LP_SOLVE` and `GLPK` can be used, additionally the `LINDO API` solver. However, some FASIMU functionality (those involving quadratic objectives and constraints) is not available with these alternative solvers.

CPLEX

CPLEX is a commercially available solver (see <http://www.ibm.com/software/integration/optimization/cplex-optimizer>). FASIMU is tested with versions 9, 10, and 12.2 but other versions should also work. IBM offers the recent version 12 free of charge for academic users, see <http://www.ibm.com/developerworks/university/academicinitiative/> for more information. There are CPLEX versions for LINUX, MacOS, Windows, Solaris, AIX, HP-IA64.

FASIMU uses the interactive solver of CPLEX which is called “`cplex`” and must be in the search `PATH`. Unfortunately, after the default installation this is normally not the case so you have to do that manually.

Locate the executable First you have to identify the location of the cplex executable which is called `cplex.exe` on windows systems and `cplex` on other systems. It can be found in a subdirectory of the main installation directory of cplex. In a LINUX installation you have set the directory manually, in the Windows installation it may have been automatically selected in a directory such as `C:\Program Files\ILOG\CPLEX`. In CPLEX 12 the subdirectory is called `cplex/bin/` followed by the name of the architecture. Be careful about correctly selecting 32 or 64 version.

Second you have to apply one of the following methods.

Setting a symbolic link You have to select a directory in the search path. The search path can be displayed with:

```
| echo $PATH
```

On LINUX I recommend using the directory `/usr/local/bin` as `/usr/bin` is tightly controlled by the installation maintenance software. If this directory is not included in the search path you can add it by including the line

```
| export PATH=$PATH:/usr/local/bin
```

in `~/.bashrc`.

Next set a symbolic link from the identified executable to the path directory. For CPLEX 12 on 64bit-Linux this may look like:

```
| ln -s /usr/local/cplex122/cplex/bin/x86-64_sles10_4.1/cplex /usr/local/bin/cplex
```

for CPLEX 10 it looks like:

```
| ln -s /usr/ilog/cplex101/bin/x86-64_RHEL3.0_3.2/cplex /usr/local/bin/cplex
```

You can also install CPLEX without super user privileges, then I recommend the directory `~/bin`. You may have to add

```
| export PATH=$PATH:~/bin
```

to `~/.bashrc` and the symbolic link command is like:

```
| ln -s /usr/local/cplex122/cplex/bin/x86-64_sles10_4.1/cplex ~/bin/cplex
```

Alternatively, extend the search path You can also add the cplex directory to the search path by adding a line to `~/.bashrc`, for example

```
| export PATH=$PATH:/usr/local/cplex122/cplex/bin/x86-64_sles10_4.1/
```

License file After the previous step is completed you should be able to call the command `cplex` in any directory. You might see the output:

```
| Failed to initialize CPLEX environment.  
| CPLEX Error 32201: ILM Error 16: CPLEX: license file not found or unreadable.  
| Exiting
```

For running CPLEX a license file `access.ilm` is required, even in the free for academics version. On many CPLEX versions also an license server demon (ilm) had to be installed but for the free version 12 this is not necessary. However, it seems to be necessary that the file `access.ilm` resides in a fixed position:

```
| /usr/ilog/ilm/access.ilm
```

for all systems except Windows

```
| C:\ILOG\ILM\access.ilm
```

for Windows. There is also a variable `ILOG_LICENSE_FILE`, which should allow to select the file position but **it does not work! Do not use his variable!** You might check out internet forums for this issue.

After the license file is in place (and the license demon process is running if necessary) you will see upon calling `cplex` something similar to this:

```
...
Welcome to IBM(R) ILOG(R) CPLEX(R) Interactive Optimizer 12.2.0.0
  with Simplex, Mixed Integer & Barrier Optimizers
...
Copyright IBM Corp. 1988, 2010.  All Rights Reserved.

Type 'help' for a list of available commands.
Type 'help' followed by a command name for more
information on commands.

CPLEX>
```

You should close this session with `^D` or the `quit` command. If that is not the case, you will have to check the correctness of the license and the installation.

LINDO API

FASIMU uses the interactive solver of LINDO API which is called “`runlindo`” and must be in the search path. If this is not already the case, there are two options to do this. Either, the directory containing the `runlindo` executable can be added to the `PATH`, e.g.:

```
| export PATH=$PATH:/usr/local/lindoapi/bin/linux64/
```

added to `.bashrc`.

Second option is to add a symbolic link in a directory in the search path, e.g.

```
| ln -s /usr/local/lindoapi/bin/linux64/runlindo /usr/local/bin/runlindo
```

You should check whether `/usr/local/bin` is in the search path:

```
| echo $PATH
```

GLPK

GLPK is an open-source free solver (see <http://www.gnu.org/software/glpk/>) which is available for LINUX and Windows. It is ported to LINUX gentoo and is probably part of other major LINUX distributions. It is ported to cygwin and can therefore the easiest way to run FASIMU under Windows. FASIMU uses its interactive solver which is called “`glpsol`” and must be in the search path which should already be the case if it is installed as part of a LINUX distribution or cygwin.

lp_solve

`lp_solve` is another open-source free solver (see <http://lpsolve.sourceforge.net/>) which is available for LINUX and Windows. It is ported to LINUX gentoo and is probably part of any major LINUX distribution. `lp_solve` is tested in the version 5.5.

FASIMU uses its interactive solver which is called “`lp_solve`” which must be in the search path. It is easiest to move the executable in `/usr/local/bin` which is the default installation. `lp_solve` requires the filter `libxli_CPLEX.so` which is included in the `lp_solve` distribution and should preferentially be placed in `/usr/local/lib`.

2.4 Install FASIMU

The installation of FASIMU itself is dead simple compared with the above. You have to place the four files `fasimu`, `fabase`, `sbml2fa`, and `reaction2fa` into a directory in the search path.

The starting point for the below commands is a running bash shell. If the current shell is not bash (see the variable `$SHELL`) problems might occur. This can easily be solved by calling

```
| bash
```

The easiest way is to install the files from the internet into `/usr/local/bin` as super user (root). You should check whether `/usr/local/bin` exists with

```
| ls -d /usr/local/bin
```

and if it does not exist you do

```
| mkdir -p /usr/local/bin
```

Then you should check whether `/usr/local/bin` is in the search path with

```
| echo $PATH
```

and if that is not the case you add

```
| export PATH=$PATH:/usr/local/bin/
```

to `~/.bashrc`. To take effect you have to start bash one more.

Then you call

```
| cd /usr/local/bin
| wget http://www.bioinformatics.org/fasimu/FASIMU.zip
| unzip FASIMU.zip
| rm FASIMU.zip
```

Alternatively, any directory such as `~/bin/` may be used to contain the FASIMU program files. This directory must also be contained in this `PATH` variable.

2.5 Visualization packages

BiNA

Download the latest version from <http://sourceforge.net/projects/binafluxmodel> which includes BiNA and the preinstalled flux analysis plugin. You need to install JAVA as a precondition. Installing BiNA is simply uncompressing the archive to any appropriate place such as `/usr/local/bina`.

Set the shell variable `BINA_HOME` to the directory the file `BiNA.jar` is located in, e.g. if bina is installed in `/usr/local/bina` add

```
| export BINA_HOME=/usr/local/bina
```

to `~/.bashrc`.

Cytoscape

Download the latest version from <http://www.cytoscape.org> and install as documented at the side. Install the plugin FluxViz as documented in <http://www.charite.de/sysbio/people/koenig/software/fluxviz/help/index.html#Installation>

CellNetAnalyzer

CellNetDesigner is a commercial network analysis tool free for academic use, see <http://www.mpi-magdeburg.mpg.de/projects/cna/cna.html>. Although graphical output is not the specific purpose of this program, a combined use with FASIMU has some advantages.

The internal data format (the files **reactions** and **metabolites**) is identical in FASIMU and CellNetAnalyzer. Thus, it is easy to transform a model developed with CellNetAnalyzer in FASIMU. The feedback of computed flux modes is easy.

Chapter 3

FABASE-models

3.1 Getting a model from CellNetAnalyzer

The format of the files `reactions` and `metabolites` equals the internal format of CellNetAnalyzer, formerly FluxAnalyzer. Some parameters in these files have no meaning inside FABASE and are retained for compatibility only. Thus, a model developed with CellNetAnalyzer is a good starting point for a FABASE model. Computed flux modes can easily be transferred to CellNetAnalyzer since the format of the `.val` files is also compatible.

3.2 Getting a model from SBML

The easiest way to produce a FABASE model is to get or produce an SBML file and use the program `sbml2fa` which comes along FASIMU. It is capable to read SBML2.1 to SBML2.4 as it only reads the reaction network which did not change in the subversions. For the metabolites it uses the tags: `id`, `name`, `compartment`, `boundaryCondition`. The latter three may be missing, after a warning a default is set. For the reactions the tags `id`, `name`, and `reversible` are being used, the latter two might be missing. Additionally, a non-standard way to encode the equilibrium constant will be read:

```
<listOfParameters>
  <parameter id="equilibrium_constant" value="...">
</listOfParameters>.
```

There are some issues where the SBML definition is not compatible with the definition of a FASIMU model. The first issue is that FASIMU encodes the compartment in a suffix to the metabolite identifier and name headed by the underscore character. Therefore underscore characters are not allowed in the compartment identifier (there are no problem in the metabolite names however). `sbml2fa` removes the underscore characters and also the space characters from the compartment names. If different compartments carry identifiers which differ only by some underscore or space characters (definitely not a sensible practice!) `sbml2fa` merges them to one. The second issue is closely related. If the metabolite identifier does not end with a `_<compartment>` already it is added. The same is done independently with metabolite names.

The second issue is that SBML allows space characters in metabolite names, but FASIMU does not (in fact, metabolite name serve as alternative identifiers in the simulation file). Therefore, `sbml2fa` replaces space characters by underscore characters in metabolite names.

Both issues result in the fact that SBMLs generated with FASIMU may have not the same identifiers as the input SBML. But if the following conditions are met, the SBML is FASIMU-proof, meaning SBMLs generated from FASIMU are compatible with the original SBML given to `sbml2fa`:

- Compartment identifiers do not contain underscore characters

- Metabolite identifiers do not contain space characters
- Metabolite identifiers end with `_` and the compartment identifier
- Metabolite names end with `_` and the compartment identifier

3.3 Getting a model from a reaction scheme

Another simple way to get a basic FABASE model is to use the program `reaction2fa` which also comes along FASIMU. It is fairly robust to read any text format reaction scheme which is available in the literature, e.g. the reaction schemes the earlier Palsson group publications uses. The basic format is a whitespace separated file where each line describes a single reaction. It starts with the reaction identifier, then the reaction using numbers (for the stoichiometric coefficients, 1 can be omitted), the identifiers of metabolites, the `+` sign and reaction arrows as `<=>` and `-->` resp. `<--` for irreversible reactions. To set the names of metabolites the option `-m <met-file>` is used. The `<met-file>` is a whitespace delimited file, the first token is the metabolite identifier (without compartment), the second token is the name which must not contain any whitespace. If reactions of the form `XXX x <=>` are found, this reaction is not transferred, instead the metabolite is marked as “open boundary”.

If there are problems with particular format, free feel to contact the author.

3.4 Writing a FABASE model from the scratch

This is easier than it might seem. For a basic FABASE model only two files are necessary, “`reactions`” and “`metabolites`” which are plain, white space separated text files. The elements surrounded by the white space (arbitrary sequences of space and tabulator characters) are called token.

Metabolites: `metabolites`

The first token is the metabolite identifier which must be unique. The next token is the name which must not contain white space. Typically, space characters are replaced with underscore characters. The next token is arbitrary for the use within FABASE. The next token is the boundary condition, 1 for metabolites which may freely enter or leave the system and 0 for all other metabolites. At this point it must be noted that the boundary condition at this point is only used for direct FABASE calculations (which are not described in this manual) and is ignored for the use in FASIMU: the boundary condition is controlled in the `simulations.txt` file.

For the use in FASIMU it is necessary to mark the compartment (consisting of letters only) as a suffix separated with a “`_`” character. Only for working with FABASE without FASIMU model they are not considered.

It is recommended to mark the names of the metabolites with the same compartment suffix. This allows to use metabolite names in the `simulations` file. If the names do not have correct suffixes the FASIMU function `simulations-work-assure` (called by `simulate`) will warn about them.

Reactions: `reactions`

The first token is the reaction identifier which must be unique. The next tokens describe the reaction, where stoichiometric coefficients are obligatory and the reaction arrow is strictly `=`. After the reaction two fixed tokens follow: “`|`” and “`#`”. The next two tokens refer to the flux bounds, the first one for the lower, the second one to the higher. “`-Inf`” is legal for the lower, “`Inf`” for the upper bound. To set the reversibility of a reaction, it must be set in the flux bound, there is no indicative reaction arrow.

3.5 Refining the FABASE Model

The following files are whitespace separated.

Equilibrium constants: `equilibriums`

In FASIMU equilibrium constants are used as weights for the backward fluxes in the flux minimization [8] and to implement the thermodynamic feasibility constraint [11]. The Gibbs free energies can not be given directly, compute the equilibrium constants with the formula:

$$K_{eq} = e^{-\frac{\Delta G_r^0}{RT}}$$

where R is the universal gas constant and T is the absolute temperature. As you can see from the formula, a FASIMU model has a fixed temperature.

Note that the equilibrium constants for FASIMU are the most frequently used dimensionless constants related to standard concentrations of 1M (not of 1mM as sometimes used).

The first item in every line of the file `equilibriums` is the reaction identifier, the second is the equilibrium constant. If the equilibrium constant of a reaction is not explicitly given it is assumed to be 1. Therefore, to exclude a reaction from thermodynamic consideration it is not sufficient not to give the equilibrium constant, it must explicitly be mentioned in `TR-exclude`, likewise in `fluxmin-exclude` to exclude it from the flux minimization scoring function.

Set fluxes to specific values: `setfluxes`

This (white-space separated) file sets fluxes to specified values. The first token of each line is the reaction identifier. If it is the only token of this line the respective flux is fixed to zero. If one token follows the flux is fixed to this value. This feature is used if the file with the name exists. Take care to delete or rename the file if it is no longer needed.

There are two differences to the file `targetfluxes` described below: (i) it is not modified by functions of FASIMU and (ii) the fluxes are always fixed to the respective values whereas there are not strictly set to the `targetfluxes` when the fitness maximization feature is used.

There is no difference to `fluxfix` when it is only used to fix it to values and not to other fluxes. The `setfluxes` overwrites the value given by `fluxfix`.

Set boundaries for fluxes: `fluxbounds`

This (white-space separated) file sets flux boundaries in conjunction to the values set in `reactions`. The first token of each line is the reaction identifier. If it is the only token of this line the respective flux is fixed to zero. If one token follows the absolute value of the flux must be lower than the value of this token. If two tokens follow they represent the lower and upper bound. “`Inf`” and “`-Inf`” are allowed, representing unrestricted fluxes.

In conjunction with other bounds is interpreted in the normal way: the maximum of all applicable lower bounds is the lower bound and the minimum of all upper bounds is the upper bound.

This feature is used if the file with the name exists. Take care to delete or rename the file if it is no longer needed.

Fix fluxes to a linear combination of others: `fluxfix`

It is possible to set that a particular reaction flux is fixed to a linear combination of other fluxes. Such reactions can also be reformulated but for clarity of the network it might be useful to retain them as separate reactions and fix them separately. An example is that 3

fluxfix is a white-space separated file. The first token is the flux to fix. The rest of the line represents the linear combination, similar to the “**enzyme**” file: First the coefficient, then the reaction identifier. The coefficient can be omitted in which case it is assumed to be one. The reaction identifier can be omitted for the last token, which means that this value is added as a direct value. Thus, the function can also be used to fix reactions to a preset value.

This feature is used if the file with the name exists. Take care to delete or rename the file if it no longer needed.

Exclude reactions from the cost function: fluxmin-excluded

In the different computations based on flux minimization every reaction is used for the objective function. Including reactions in “**fluxmin-excluded**” is an easy way to exclude certain reactions (e. g. those which do not describe a cellular effort).

Set relative weights for the cost function: fluxmin-weights

By default, every flux has the same relative weight in the scoring functions in the flux minimization principle. The file **fluxmin-weights** however, changes the relative impact of individual functions. Not all reactions have to be mentioned in this file, in this case 1 is assumed. Also the weight 0 can be given, it has the same effect as including the reaction in **fluxmin-excluded**. The function **fluxmin-excluded** takes precedence over **fluxmin-weights**, that means if a nonzero weight is given in **fluxmin-weights** but is also mentioned in **fluxmin-excluded** then the reaction is excluded from the score calculation.

Concentration ranges

Concentration ranges must be supplied if TR-modes should be computed. To simplify specification of concentration ranges, concentrations may be given for classes of metabolites rather than for metabolites directly. The file **concentration-ranges-classes** assigns a metabolite to a class.

The file **concentration-class-ranges** then assigns concentration ranges to classes. Each line usually contains 6 tokens: The class identifier, the lower hard bound, the lower soft bound, the set point, the upper soft bound, and the upper hard bound. The hard bounds must never be exceeded. The soft bounds may be violated at the expense of an additional penalty. If a concentration value differs from its set point, a (usually low) penalty is given in the objective function (if this function is switched on). Depending on the number of tokens the following rules apply:

Number of values	Ranges definition	Applied assignments
5	<low hard> <low soft> <setpoint> <high soft> <high hard>	
4	<low hard> <low soft> <high soft> <high hard>	<setpoint>= $\sqrt{\text{lowsoft} \times \text{highsoft}}$
3	<low hard> <setpoint> <high hard>	<low soft>=<low hard>, <high soft>=<high hard>
2	<low hard> <high hard>	<setpoint>= $\sqrt{\text{lowsoft} \times \text{highsoft}}$, <low soft>=<low hard>, <high soft>=<high hard>
1	<fixed concentration>	

For most applications (when no concentration prediction is required) only the hard bounds are required. For the soft bounds and the set points there is also allowed to write a minus sign instead of a value. This is interpreted as there is no boundary for this particular metabolite class. Also it is possible to assign a higher importance to particular set points or soft bounds:

| <value>(<weight>)

where the default weight is 1.

Exclude reactions from TR-computations: TR-exclude

Thermodynamic feasibility can only be assessed if the standard Gibbs free energies of a reaction is known. Therefore FASIMU/FABASE allows to selectively exclude reactions from the TR constraint: put the identifier of the reaction in the file **TR-exclude**. Often, Gibbs energies of reactions are computed as the difference of formation energies of products and reactants. If a formation energy is not known, the Gibbs energy of all reactions involving this metabolite is presumably also not known. Thus, another option is to enter a metabolite identifier in **TR-exclude**: this means that any reaction involving this metabolite is excluded from the TR criterion. Note that the identifier must include the compartment suffix. This is a consequence that TR resides at the FABASE-level which (as opposed to FASIMU) does not know about compartments. **TR-exclude** is a white-space separated file, the tokens are accepted no matter if they appear in the same line or in different lines.

Additional names of reactions: reaction-names

This is a white-space separated file: the first token on each line must be a reaction identifier, the rest of the line is regarded as the name of the reaction. The name of the reaction is for instance added at the end of each reaction flux line in **allout.txt**.

Enzymes and specific weights: enzymes

FASIMU implements enzyme cost minimization. The information how the reaction fluxes are related to enzymes are defined in the file **"enzymes"**. The syntax is as follows. It is a white-space separated file. The first token on each line is the identifier of the enzyme. It is allowed that identifier also appears as a reaction identifier, enzymes and reactions have different name spaces. Then, a coefficient follows, then a reaction identifier. The meaning of the combination $E \dots cR$ is that the enzyme E requires the cost c to catalyze a unit flux through the reaction R . The coefficient can also be omitted, in which case it is considered to be 1. The coefficient zero is also allowed, it means that the respective reaction is assigned no cost with respect to this enzyme. For the enzyme cost minimization this has no effect but it records the relation of the reaction to the enzyme.

Several important notes for this function:

- As opposed to other files in the FA-model it is not automatically used, the respective optimization function must require this: **compute-FBA -e**.
- A reaction which is not referred to in **"enzymes"** is considered to have no cost.
- The information is only used for the enzyme minimization algorithm.
- The use of this file requires that the reaction identifiers may must not consist only of digits, it is clear that they can be confused with the coefficients.

FABASE files used to implement FASIMU: targetfluxes and fluxconstraints

The file **"targetfluxes"** provides a way to fix the flux rate of a reaction (first token) to a value (second token). The file **"fluxconstraints"** borders the flux value. An upper limit is given as one token, whereas a range requires two tokens. The simulation objective (see **simulations** below) is implemented with **targetfluxes**, the simulation constraints are implemented with **fluxconstraints**. Since FASIMU automatically generates these files prior invocation of a FABASE function, these options are only available when FABASE is used without FASIMU (see below).

3.6 FASIMU-compatible Models

For compatibility, conventions of CellNetAnalyzer have been adopted for FABASE and FASIMU:

- Space characters are not allowed in identifiers.
- Reaction identifiers may be not by number number.
- Every metabolite must have an assigned compartment where the compartment is attached as suffix to the metabolite identifier preceded by underscore.
- Compartment identifiers must consist only of letters.
- The compartment “**ex**” is not allowed.

3.7 Modify model files in a FASIMU session

The recommended way to handle your models

Generally, I do not recommend to modify the files in the directory in which you are working with FASIMU. The program may be capable to regenerate intermediate files and for advanced users it makes sense to do so (see below). But to start with I strongly recommend that you keep the original models in separate directories and start fasimu in a fresh directory. Say, you keep and modify your model in a directory `~/model` then proceed like that:

```
cd ~
cp -R model model-compute
cd model-compute
source fasimu
<do the computations>
<copy the result files to a designated directory>
cd ..
rm -r model-compute
```

If you are acquainted with FASIMU you will certainly design your own work flow.

Modification functions

If you choose to modify the model files you must not change the files `reactions`, `metabolites`, `equilibriums`, `TR-excluded`, `fluxmin-excluded`, `concentration-ranges-classes` but the files with the extension `.original` instead. After the modification you call `source fasimu`. You may change the file `simulations` any time you wish. The changes come into effect the next time you call `simulate` or `simulate-single`. For other files you have to call update functions as follows:

Modified file	possible	Function for the safe integration of the modification
reactions	no	change reactions.original instead
reactions.original	yes	fasimu-main
metabolites	no	change metabolites.original instead
metabolites.original	yes	fasimu-main
equilibriums	no	change equilibriums.original instead
equilibriums.original	yes	fasimu-main
TR-excluded	no	change TR-excluded.original instead
TR-excluded.original	yes	fasimu-main
fluxmin-excluded	no	change fluxmin-excluded.original instead
fluxmin-excluded.original	yes	fasimu-main
concentration-ranges-classes	no	change concentration-ranges-classes.original instead
concentration-ranges-classes.original	yes	fasimu-main
enzymes	yes	update-enzymes
fluxmin-weights	yes	not necessary
fluxfix	yes	update-fluxfix
fluxbounds	yes	update-fluxbounds
setfluxes	yes	update-fluxbounds
concentration-class-ranges	yes	update-concentration-ranges
simulations	yes	not necessary
constraint file referenced within	yes	touch simulations
expressions	yes	not necessary
cplex-tail.in	yes	not necessary
lp_solve.par	yes	not necessary
lindo.par	yes	not necessary
\$glpk_opts	yes	not necessary
targetfluxes	no	not to be used in a FASIMU session
fluxconstraints	no	not to be used in a FASIMU session

Chapter 4

The steps in a FASIMU session

4.1 Starting FASIMU

As mentioned fasimu is invoked by

```
| source fasimu
```

There are some options to this call which are indicated by the following identifiers which follow the word fasimu in the command line.

cplex, **cplex9**, **cplex10**, **lp_solve**, **glpk** selects the specific solver. This is necessary if the automatic selection of the solver fails or selects not the desired solver. The automatic recognition of the version of cplex calls **cplex** but that might fail if this is a single license installation and another user is using it at the moment. In this case **cplex9** is used for versions up to 9 and **cplex10** for version from 10 on.

names_in_allout. Normally the identifiers of the metabolites are written in the file allout.txt which also. If you prefer the names instead, select this option.

nomodelprepare Here, only the FASIMU functions are defined but any action on the model is omitted. This is useful if only a file allout.txt or evaluation.txt from a previous run shall be analyzed or used with the FASIMU functions, for instance for the visualization.

debug Some debug messages in the course of the fasimu invocation.

4.2 The control file simulations

The general structure is as follows:

- Each line describes a single simulation
- Each line contains four tab-separated fields: name, objective, constraint, evaluator, comment. Each of the fields may contain several tokens, separated by the space character.

First column: Name

That is the primary key and must be a unique identifier, imulations are identified by its name. It may contain space characters but the character / is removed.

Second field: Targetflux(es),Objective(s)

The second field contains targetfluxes and mimimization or maximization objectives. Targetfluxes are fluxes that are set to a specific value (or in a fitness maximization optimization, fluxes that should come as close

as possible to this value). After the keyword **max** fluxes are marked which should be as high as possible, after the keyword **min** as low as possible (including negative values if marked as reversible), and after the keyword **minabs** whose absolute value is as low as possible.

The field is organized as a separated list of tokens. A token is either a reaction-ID, a metabolite-ID, or a decompartmentalized metabolite-ID (in this case it is defaulted to the compartment indicated by the variable **default_compartment** which is **ext** by default). Each token can be preceded by a real valued coefficient ("1" is the default). The coefficient has two different meanings: for the targetfluxes it is the *flux value* the flux is set to, while for the min/max objectives it is the *weight* in the scoring functions. This weight is relevant if several fluxes are marked for max/minimization, but also if more items are combined which modify the objective functions, such as fitness maximization terms, as well as using set points or soft bounds in thermodynamic realizability calculations. Negative weights are not recommended¹

Several **min**, **minabs**, or **max** sections can be combined but targetfluxes can only be noted before the first **min**, **minabs**, or **max** keyword. As opposed to the third column, tokens in this field are used without qualifiers

For min/maximization problems the user has to take care that the system is restricted to reasonable flux ranges. In fact, FASIMU sets bounds of $-10^6, 10^6$ to the marked fluxes to circumvent a unbounded optimisation problem. The two most frequent applications are

- the maximization of biomass or a metabolite while restricting the inflow of substrates. This restriction can conveniently be defined in the third field.
- the substrate intake minimization while a production rate of biomass or some metabolites is set as a targetflux. Here, it is important to set in the intake reaction as irreversible.

Third column: System boundary tokens

Simplified syntax

Each token is headed by -, +, =, or %, followed by an identifier as above.

	Reaction	Comment	Metabolite	Comment
+	forward direction	definition of forward/ backward direction: see below	product	export of this metabolite across the system boundary is allowed
-	backward direction	definition of forward/ backward direction: see below	substrate	import of this metabolite across the system boundary is allowed
=	not applicable	allowing non-zero flux through reactions is the default	product or substrate	import or export across the system boundary are allowed
%	forbidden	flux through reactions must be zero	not applicable	zero flux through exchange reaction across the system boundary is the default

The rules dictated by the above constraints are combined with flux bounds given in the file **reactions**.

Explicit boundary syntax

Another syntax is possible in the constraints section:

Syntax	Comment
<identifier> = <number>	Fixed flux for the respective reaction or system exchange
<identifier> <= <number>	Upper bound for the respective reaction or system exchange
<number> <= <identifier>	Lower bound for the respective reaction or system exchange
<identifier> >= <number>	Lower bound for the respective reaction or system exchange
<number> >= <identifier>	Upper bound for the respective reaction or system exchange
<number> <= <identifier> <= <number>	Range for the respective reaction or system exchange

¹In fact, a negative weight to a **min** has the same effect as using the inverted weight to a **max** section and vice versa. A negative weight to a **minabs** does not work.

There must a space characters surrounding the \leq or $=$ character(s). Instead of \leq also $<$ can be written but the result is identical because in the optimization software (and in fact for the underlying theory of optimization) strictly “less than” is not implemented.

File content syntax

In this column file-content-replacement may avoid repeating long lists several times: When the name of a text file appears in this column, its name is replaced by the file content.

```
| .. stddef ...
```

where the file `stddef.txt` contains the lines

```
| Glc_ext
| 0 <= PFK <= 2
| +C02_ext
```

is equivalent to the constraint definition

```
| .. -Glc_ext 0 <= PFK <= 2 +C02_ext ...
```

Precendence/Overwrite

If there is more than one token constraining the same reaction, only the rightmost constraint is valid, the constraints further to the left are ignored. In other words, the list of constraints is evaluated from left to right and the constraints regarding the same reaction are overwritten. Note that they are not combined: the constraints

```
| -Glc_ext ... -8 <= Glc_ext <= 8
```

are interpreted as the glucose exchange flux between (-8,8), not (-8,0).

This may be confusing if one of the constraints is contained in a file. Say, oxygene uptake is allowed in the file `stddef.txt` with a line `-O2_ext` and you want to model an anoxic state. You must write

```
| stddef ... %O2_ext
```

If you write

```
| %O2_ext ... stddef
```

you the token `%O2` will have no effect as it is overwritten by the setting in the file `stddef.txt`.

Fourth column: evaluator

This is used to ensure a quick overview whether a simulation is successful (indicated by a $+$ sign in the second column of `evaluation.txt`). It can be either zero (this means this simulation is intended to fail), one (the existence of a solution suffices), or one (or several) token(s) as above in which case it is checked whether the reaction associated with this token carries a nonzero flux in the flux solution. A metabolite ID can be

- its model identifier with or without compartment
- metabolite name with or without compartment

If more than one evaluator is given the simulation is considered to be satisfied if all evaluators are satisfied. However, the comment in column 3 of `evaluation.txt` indicates which of the tokens are tested successfully.

Fifth column: comment

This is technically not used in FASIMU but it is a good idea to describe the simulation semantically here.

4.3 Controlling the computation function

There is one function for all sorts of flux optimization called `compute-FBA`. The variable `$optimization_call` should be set to one of the functions below together with parameters, e.g. `optimization_call=compute-FBA -T A -F e -s 1 -w 0.0001''`.

Computing an arbitrary solution: `compute-FBA`

Without any arguments, a solution is computed which satisfies the given constraints (including the flux-balance condition). Which solution is returned is basically left to solver as *no scoring function is assumed*. Typically, the solver returns a solution with as few as possible non-zero fluxes but you can not rely on that indeed the flux solution with the absolute minimum is given (you have to set `-F m`) to do this. The advantage is that the computation is the fastest. It is the recommended way to compute an FBA solution just to check the feasibility of the constraints.

The flux minimization: `compute-FBA -F`

A flux distribution obeying the flux-balance condition is computed. It minimizes the fluxes as a general rule, but there are some variants, controlled by the single parameter. The default is an implementation of [8] where the forward fluxes have the weight 1 and the backward fluxes have the weight equal to the equilibrium constant (given in the file `equilibriums`). If the parameter `-F s` is given the weights of the forward and backwards flux are divided by $\sqrt{1 + K_{eq}^2}$, according to [10]. The parameter setting `-F 1` follows the same idea, both weights are divided by $1 + K_{eq}$. For the parameter setting `-F 2`, the forward flux has the weight $1/(1 + K_{eq}^2)$, the backward flux the weight $K_{eq}^2/(1 + K_{eq}^2)$. For the parameter `-F 0`, the backward flux has the same weight as the forward flux, regardless of the value in equilibriums. This is equivalent to the case where no equilibrium data is given at all. For the two options `-F e` and `-F E` the file enzymes must be available. Here, weights are not given for the fluxes themselves but for the enzyme with respect to a reaction. The general principle is also given in [9]. The difference between `e` and `E` is that for `e` both directions of the reaction receive the same weight where for `E` the backward flux is multiplied with equilibrium constant, similar to the default.

The call `compute -F m` minimizes the number of nonzero fluxes. If there are no other constraints than system boundary exchanges then the computed flux distribution is an **Elementary flux mode** [19].

Allowing restricted fitness: `compute-FBA -f`

This implements the generalized flux minimization principle [9]. Fluxes must not strictly obey the given objective (in FASIMU, the objective tokens in `simulations`) but are allowed to deviate. Thus, the optimization maximizes the fitness as the first priority.

The fitness function ranges from zero to one, where one represents maximal fitness. This figure is printed in the output files.

The option `-f` takes 2 parameters. The first one is the type of the fitness function, ranging from 0 to 7. Default is 0. For solvers LINDO, `lp_solve` and GLPK quadratic scoring functions are not implemented, so the type may only range from 3 to 7, and the default is 5. The types are distinguished by the way the aberration to the target flux is combined: by the Euklidian norm (0,1,2), by the 1-norm (3,4,5), by the maximum norm (6,7). They are also distinguished whether the distances are normalized or not, i.e. if absolute or relative distances are considered. Type 1 is special as it involves a partial normalization. Type 3 is a very easy and fast implementation but for this it is necessary that the flux values have the same sign as the target value

for the target fluxes (it is guaranteed in the computation). Type 8 is similar to type 2, just the square root is omitted in the scoring function. The computed flux solutions are identical.

Type	Norm	Adjustment	Fitness score	Restriction
0	Euklidian	none	$1 - \sqrt{\frac{\sum \Delta v_i^2}{\sum L_i^2}}$	$\text{sgn}(v_i) = \text{sgn}(L_i)$
1	Euklidian	partly (linear)	$1 - \sqrt{\frac{\sum \frac{\Delta v_i^2}{ L_i }}{\sum L_i }}$	
2	Euklidian	full	$1 - \sqrt{\frac{1}{n} \sum \frac{\Delta v_i^2}{L_i^2}}$	
3	linear	full	$1 - \frac{1}{n} \sum \frac{ \Delta v_i }{ L_i }$	
4	linear	none	$1 - \frac{\sum \Delta v_i }{\sum L_i }$	
5	linear	full	$1 - \frac{1}{n} \sum \frac{ \Delta v_i }{ L_i }$	
6	maximum	none	$1 - \frac{\max\{ \Delta v_i \}}{\max\{ L_i \}}$	
7	maximum	full	$1 - \max\left\{\frac{ \Delta v_i }{ L_i }\right\}$	
8	quadratic	full	$1 - \frac{1}{n} \sum \frac{\Delta v_i^2}{L_i^2}$	

L_i is the target flux value. $\Delta v_i = v_i - L_i$ is the distance of the actual flux value to the target flux value. The sums and sets are over the target reactions i .

The second parameter is the weight with respect to the weight of the fluxes. Technically, the optimizer does not supply a hierarchic optimization, thus, a large weight has to be given to the fitness score. The default is 10^6 .

Fitness maximization is closely related to MOMA (minimal metabolic adjustment) and can in fact implemented by defining the targetfluxes according to a reference solution. However there is also a special FABASE function `compute-moma` for this purpose.

Minimizing a selected target: `compute-FBA -m`

This function allows to minimize one or more reaction fluxes, usually applied to the inward transport process of substrates. The parameter to this option is one or more reaction identifiers.

There are two ways to use this option in FASIMU. In the first one `-m` is included in `$optimization_call`. In the `simulations` file the objective (second column) is left blank (or used to fix some other fluxes). The disadvantage of this way is that for all simulations the minimization of the same reaction/metabolite is performed.

The second, strongly recommended, way does not have this restriction. The variable `$optimization_call` contains a `compute-FBA` call without the `-m` option but possibly with other parameters. In the `simulations` file the objective (second column) is `min <rea/met> ...` with one or more tokens (reaction identifier, metabolite identifier or names). The subroutine of `simulate` executing this simulation automatically adds `-m <reaction-ID>` to `$optimization_call`. Obviously, this only works if the `$optimization_call` is `compute-FBA` with possible further parameters.

Maximizing a selected target: `compute-FBA -b`

This option allows to maximize one or more reaction fluxes, usually applied to the biomass synthesis process or an important product [4]. The parameters to this function are the reaction identifiers.

Here the same mechanisms can be used as in the previous section. The respective keyword is `max` instead of `min`.

Thermodynamic feasibility computations: `compute-FBA -T`

This option switches on the criterion that a positive flux through a reaction must be accompanied by a negative Gibb's free energy the latter being dependent on standard Gibb's energies and ranges of allowable

concentrations. The so-called criterion of thermodynamic realizability (TR) is described in detail in [11]. This option requires equilibrium and concentration ranges to be set (see above).

TR is only a constraint, the flux objective is set by the above options `-F`, `-m`, and `-b`, with the default of `-F 0` if just called with `compute-FBA -T`.

The TR computation not only predicts fluxes but also metabolite concentrations, stored in the file `concentrations.txt`. With the option `-d` given to `compute-FBA` a more comprehensive report on concentrations is stored in the file `TR-FBA-concentration-doc.txt` comprising also the applied ranges. `-d` gives also further information files: `TR-FBA-potential-doc.txt`, `TR-FBA-potential-doc2.txt`, and `TR-FBA-summary.txt`. Potentials (see below) are the sums of logarithms of the reactants plus the logarithm of the equilibrium constant. The documentaries on potentials show it for individual reactions. The summary shows the whole model together with the flux distribution and the concentrations.

For good concentration predictions reasonable soft bounds (switched on with the `-s [<weight>]` option) and set points (switched on with `-w [<weight>]`) should be supplied for the concentrations. Typical call is `compute-FBA -T -s 1 -w 0000.1`. The default weight for the soft bounds is 1, the default weight for the setpoints is 10^{-5} , so this call is equivalent to `compute-FBA -T -s -w`. Note that it is not enough that the soft bounds and the setpoints are defined in `concentration-class-ranges`, they are only used if `-s` or `-w`, respectively, are given.

For the TR criterion a type identifier can be given which controls the way, zero fluxes and thermodynamic potentials are treated. Basically, a non-zero flux and a non-zero potential must have the same sign. (Thermodynamic potential here is the negative Gibb’s energy change divided by RT .) But it is not clearly defined if one of the values is zero. There are two ways: in the relaxed form (a or A) a zero potential is compatible with any flux value, and a zero flux value is compatible with any potential. In the strict form (b or B) a zero potential is only compatible with a zero potential and vice-versa. The differences in the biochemical interpretation will not be covered here. But from the computational side: types b and B are much harder to compute (on more binary variable for each reaction) and the system is much more likely to be infeasible. The uppercase letters refer to an implementation of TR with conditional clauses which is more robust, numerically stable, and accurate in the computation but is only available in the solver CPLEX version 10 and higher. The lowercase letters refer to the so-called bigM implementation which is the standard for other solvers.

A trailing “r” in the type identifier changes the way reaction marked as irreversible are dealt with. By default, there is still the binary variable. For a zero flux, the potential can have either sign. By the setting “r” there is no conditional variable but the sign of the potential is fixed to the sign of the irreversibility constraint. If the flux is non-zero, that is not a difference. However, for a zero flux, the constraint on the potential is still active, where in the default way, there is also a potential with the opposite sign possible. Again, the biochemical implication will not be discussed here. From the computational side, the setting “r” accelerates the computation but also increases the likelihood that the system is unfeasible.

Type	Zero flux	Zero potential	Irreversible reaction	comment
A	q arbitrary	any flux		Conditional clause implementation
B	$q = 0$	$v = 0$		Conditional clause implementation
a	q arbitrary	any flux		bigM implementation
b	$-\epsilon < q < \epsilon$	$-\epsilon < v < \epsilon$		bigM implementation
Ar	q arbitrary	any flux	potential sign fixed	Conditional clause implementation
Br	$q = 0$	$v = 0$	potential sign fixed	Conditional clause implementation
ar	q arbitrary	any flux	potential sign fixed	bigM implementation
br	$-\epsilon < q < \epsilon$	$-\epsilon < v < \epsilon$	potential sign fixed	bigM implementation

See the section about concentration ranges above as they are necessary for the TR computation.

Calculating relative fluxes with compute-FBA

The flux bounds (and more specifically the irreversibility of reactions) in the stoichiometric network model (represented by the file `reactions`) have the purpose to restrict the absolute reaction flux rates in the system.

To calculate flux changes (relative fluxes) they are not valid. For example, the reaction i may be considered irreversible, i.e. $v_i \geq 0$, but the flux may change in negative direction: $\delta v_i = v_i - \hat{v}_i$ may be negative if v_i is smaller than \hat{v}_i .

FASIMU offers two mechanisms to compute relative flux distributions without modifying the model, one for the situation where a reference flux distribution is given and the other one if the reference flux distribution is not given.

With reference flux distribution

```
| optimization_call="compute-FBA -r solution.txt ... "
```

Technically, the values of the reference flux distribution are subtracted from the bounds given in **reactions**. Note, that the values in the file **fluxbounds** are not affected because they are not considered to be part of the network — their purpose is to affect a single calculation. The same is valid for the boundary expressions in the file **simulations**.

Without reference flux distribution

```
| optimization_call="compute-FBA -delta ... "
```

Without a reference there is no basis to have specific upper and lower bounds, so the effect of this parameter is simply to allow negative values for irreversible reactions. The implementation generalizes this to be also applicable for bounds which are not **-Inf**, **0**, or **Inf**. Technically, the maximum m_i of the upper bound and the negative of the lower bound is computed and the flux is bounded by $-m_i \leq v_i \leq m_i$.

Further options of compute-FBA

There is another parameter worth to mention: **-t <seconds>** which restricts the computation time of a single optimization.

MOMA: compute-moma

This function implements MOMA [20] using the fitness function (see above). The original paper uses unnormalized Euklidian distance but that can be changed by setting the **-f** option. In fact, all options of compute-FBA can also be given to compute-moma. The Euklidian distance can only be used if the solver is CPLEX, otherwise it is switched to the non-normalized linear type (**-f 5**). **compute-moma** has its own help text available with **compute-moma -h**.

ROOM: compute-room

This function implements Regulatory on/off minimization [21]. As opposed to compute-moma it is a separate implementation and has only a few parameters to be displayed with **compute-room -h**. It is possible to change the relative and absolute threshold (δ and ϵ in the original paper are set by **-d** and **-e** respectively).

Expression-based flux prediction

Binary use of expression profiles: compute-FBA -xs

The option **-xs** of **compute-FBA** implements the algorithm to predict the active subnetwork depending on a expression profile (which can be a protein or transcript profile) [22]. The profile is given in the file **expressions**, which is white-space separated. The first token is the reaction identifier, the second is an expression value, which can be a float number, a binary or a three valued measure. The option can have up to four parameters:

Parameter	Description	Default
1st	lower threshold of expression	0.5
2nd	upper threshold of expression	equal to lower threshold
3rd	threshold on significant flux	1
4th	relative weight compared to other scoring components	1000

The interpretation of the expression profiles is controlled by the lower and upper threshold. Transcript values below the lower threshold are considered *off*, a flux through the respective reaction is penalized (with 1). Transcript values above the upper threshold are considered *on*, a significant flux (3rd parameter) through the respective reaction receives a bonus. Expression values between the thresholds neither are penalized nor receive a bonus (of 1). Reactions not referred to in the file **expressions** are also excluded from the scoring.

The function `compute-shlomi` is a standalone version of this function. See also the help available with `compute-shlomi -h`.

Gradual penalty to *off* expression: GIMME `compute-FBA -xg`

The option `-xg` of `compute-FBA` implements another algorithm to predict the active subnetwork depending on an expression profile (which can be a protein or transcript profile) [2]. Again the file **expressions** is used in the format as above. The option can have up to two parameters:

Parameter	Description	Default
1st	threshold of expression	0.5
2nd	relative weight compared to other scoring components	1000

Expression values x_i below the given threshold t receive a penalty of $|v_i|(t - x_i)$ where v_i is the respective flux rate belonging to the expression value. There is neither a bonus nor a penalty for expression values above the threshold. Obviously, the penalty is zero if the respective flux rate is zero.

Combined scoring function

In `compute-FBA` any expression-based flux prediction score can be combined with other factors. One example is the combination with flux minimization [8] and fixing metabolic target functions (enforce the production of biomass):

```
| optimization_call="compute-FBA -xs -F"
```

See [12] for details on the method, see tutorial for an example. The scoring function can also be combined with fitness minimization (-f), intake minimization (-m), biomass maximization (-b). Even the binary expression scoring can be combined with GIMME scoring, the respective weights and threshold act independently.

4.4 Perform simulations: `simulate`

The function call `simulate` performs all simulations described in **simulations** and stores the complete result in **allout.txt** and a short overview in **evaluation.txt**. To just compute a single simulation identified by its name call `simulate-single "<name>"` where the quotes can be omitted if the name contains no space characters. To store the solutions requires considerably disk space for large networks, `simulate-noallout` performs the computations but suppresses to write the solutions.

The call of `simulate` deletes the files **allout.txt** and **evaluation.txt** as the first step, thus, it is guaranteed that after a `simulate` call those files contain the information on one complete set of simulations. The call `simulate-single` however appends the result to **allout.txt** and **evaluation.txt**. This way they may contain several solutions of the same simulation.

The call `compute-FBA` stores information which is overwritten by the next call which is the case if `simulate` is called and there is more than one simulation. If you are interested in this information, call `simulate-single` instead and save the information in separate files. See the following table for the information:

Overwritten information	
<code>problem.lp</code>	complete optimization problem description
<code>solver.out</code>	solver output
<code>variables.out</code>	values of all variables
Appended information	
<code>evaluation.txt</code>	summaries
<code>allout.txt</code>	solutions
<code>cplex-times</code>	Running times (CPLEX only)

The function `simulate` allows one or two parameters which are numbers of simulations (not names! The numbers are not written in the `simulations` file.) `simulate i` executes the simulation with the number `i`. `simulate i j` executes all simulations starting with number `i` until number `j`. If `j` is smaller than `i` no simulation is performed.

4.5 The evaluation file: `evaluation.txt`

This file is tab separated:

1. Name of the simulation
2. + or - depending if the computed result is in accord with the expectation
3. A comment on the evaluation

There are several bash functions which provide convenient views on the result:

`negeval` gives only lines in `evaluation.txt` which have not the desired result.

`inacceval` scans the file for warnings on the inaccuracy of results (switched on by the `-c` option of `compute-FBA`).

4.6 The comprehensive solution file: `allout.txt`

The file contains all information on the solutions: flux values, concentration values if applicable in a human readable form. Each solution begins with a line with many `\#` characters and the name of the simulation. Then a message on computation and possible warnings follow. After that the reaction fluxes follow in a tab separated format, the first item is the reaction identifier, the second item is the flux rate, the third is the equilibrium constant used, the fourth is the reaction equation, the fifth are possible annotations of the reactions (given in reaction-names). The reactions are normally written with the identifier of the metabolites. However, it can also be written using the names of the identifier by invoking `fasimu` with:

```
| source fasimu names_in_allout
```

Zero fluxes are omitted throughout, although occasionally fluxes with flux rates close to zero may appear in large networks which are numerically difficult. The first fluxes denoted in the solution are the fluxes across the systems boundary (which are not fluxes in the metabolic reaction systems), identifiable by the trailing “`_tr`” in the reaction identifier and the trailing “`_ex`” in the metabolite identifier in the reaction equation. Then the regular fluxes follow. If applicable, i.e. if `compute-FBA -T` was used, concentrations of the metabolites involved in the solution follow, in a tab-separated format: the first item is the metabolite identifier, the second is the concentration value, and the third are the concentration ranges used in the computations which might include soft bounds and set points if they have been used.

4.7 Prepare files for the visualization

FASIMU itself has no visualization capabilities on its own, however it includes plugins for the easy integration of computed flux modes in other packages.

Visualization in Cytoscape

For the visualization in [14] in combination with the flux analysis plugin, the call of `allout2valfiles` transforms the solutions recorded in `allout.txt` in separate files in a directory `val`.

Visualization in BiNA

For the visualization in BiNA [16] in combination with the flux analysis plugin (<http://binafluxmodel.sourceforge.net/>), the call of `allout2bina` transforms the solutions recorded in `allout.txt` in separate files in a directory `BiNA`. The names of the files are the respective simulation names. There are two parameters to this function. The metabolites (without a compartment identifier) following “-a” are defined to be alias metabolites (see BiNA documentation), metabolites which are not drawn as one node in the graph but as many nodes possible distributed throughout the graph. E.g.

```
| allout2bina -a ATP ADP NADH NAD
```

defines these metabolites to be aliased in all occurrences. BiNA is capable to define alias function for each reaction separately but this has to be modified manually, see the description of the BiNA-flux analysis file format. The parameter `-c` defines RGB color values for the color of metabolite nodes depending on the compartment. The syntax is as follows: `compartmenti[numRi,numGi,numBi]`. This sequence must be written without space characters. The respective values range from 0 to 255. In the `-c` option an arbitrary number of such items may be given. Example:

```
| allout2bina -c cyto[245,45,226] mito[66,2.3.344]
```

Note that if all three numbers are low the color is very dark and the black letters of the metabolite name may not be readable.

A previous directory `BiNA` will be removed by this function. If the file `allout.txt` contains more than one flux distribution with the same name (this happens if the same simulation is run over again with `simulate-single`), a underscore character and a number (starting from 0) is appended to the name of the simulation to avoid overwriting.

Visualization in CellNetDesigner

For the visualization in [15], the files stored in the `val` directory by `allout2valfiles` can also be directly be used for the visualization in CellNetDesigner by the `ReadMode` function. Of course, CellNetDesigner requires a ready-made image of the network.

Chapter 5

Further functions in FASIMU

5.1 Prune to *ad functionem* networks: `prune-network`

The function checks for each reaction whether a positive or negative flux is possible. This depends on the selection of the optimization protocol (variable `$optimization_call`) and the contents of the file `stdexch.txt` which defines the system boundary. See [7] for details on the method. Both, the exchange metabolites and the functional metabolites mentioned in the article must be included in the same file `stdexch.txt`.

The result of the function is a new network in the subdirectory `sub`. The blocked reactions are removed from the network. If only one direction is blocked it is marked as irreversible in the newly created file `sub/reactions`. The resulting files are a valid FAFASE model, i.e. it does not contain the FASIMU pseudo reactions controlling the systems boundary.

If an error message “`bash: declare: -A: invalid option`” is shown, you should update your bash to a version higher than 4.

If that is not possible you can use the function `prune-network-simple` which should obtain the same result but is much slower as doesn't use already computed flux solutions to reduce the number of required optimization objectives.

5.2 Flux-variability analysis

The function `make-FVA-simulationsfile` prints a simulations file applicable to perform a flux-variability analysis [17,18] which should be directed to `simulations`. There are two other elements to be taken care for a meaningful FVA. The first is the file with the predefined name `stdexch.txt` which defines the constraints valid in all defined simulations and it should be used to allow input of substrates and output of waste products and metabolic objectives. The second is the definition of flux boundaries. If there would be no flux restrictions probably many of the fluxes were unbounded in which case the FVA would be meaningless. It is recommended to either restrict all input fluxes or all output fluxes.

The function `FVA-valfiles-chart` interprets the result files of a FVA to produce a chart, list of reactions with their respective minimum and maximum. It uses the files in the directory `val` thus, it is necessary to call `allout2valfiles` beforehand. See tutorial for the normal work flow:

```
make-FVA-simulationsfile > simulations
simulate
allout2valfiles
FVA-valfiles-chart
```

This version of FVA directly maximizes and minimizes the problem. If there are problems that the optimization problem is not bounded, you can use another form of FVA which only checks whether it is

possible to find a flux solution with a unit flux in both directions for each reaction:

```
make-unity-FVA-simulationsfile > simulations
simulate
allout2valfiles
FVA-valfiles-chart
```

5.3 Flux control analysis

The function `make-FCA-simulationsfile` prints a simulations file applicable to perform a flux coupling analysis [3] which should be directed to `simulations`. The file with the predefined name `stdexch.txt` must define the constraints valid in all defined simulations and it should be used to allow input of substrates and output of waste products and metabolic objectives.

Before you start make sure that there are no blocked reactions (perform pruning first) as these reactions yield misleading results.

The function `FCA-chart` interprets the result files of a FCA to produce a chart. It is a tab-separated file with four columns:

1. analyzed reaction
2. statement on coupling, on of
 - fully coupled to
 - uncoupled with
 - directionally coupled to
 - partially coupled to
3. reference reaction
4. additional comment (such as `forbidden condition`)

The protocol to perform the analysis (after pruning) is:

```
make-FCA-simulationsfile > simulations
simulate
FCA-chart
```

Note that in FASIMU reactions in both directions are allowed while in the original manuscript [3] a framework is assumed which treats the directions as separate reaction entries. Thus, the respective simulations are marked with `forward` and `backwards`.

5.4 Check the essentiality for a given set of simulations: `check-essentiality`

This functions checks the essentiality of reactions with respect to the defined simulations. First, the actual simulations (defined by the `simulations` file and the variable `optimization_call`) are performed. Based on the results a new set of simulations is defined implementing a single knock-out for each reaction contained in the solution of the respective. Note that this is much more efficient than to check every reaction — a reaction can only be essential if it appears in a reference solution.

The result is written concisely in `essentiality-report.txt`. Each line refers to a simulation in the original set. The lines are tab-separated, the first token is the identifier of the simulation and the second is either

- the comment `failed` if the original simulation failed
- the comment `no essentials` if they are no essential reactions, or
- `essentials`: followed by a space separated list of reaction identifiers referring to the essential reactions.

Other intermediate files can also be inspected:

- `evaluation.txt` contains the result of the reference run,
- `simulations_essentiality` contains the newly defined simulations,
- `evaluation_essentiality.txt` contains their results.

5.5 Functions creating common simulations files

The following functions produce simple `simulations` files for common applications.

Check the producibility of all metabolites

```
| make-fullproducibility-simulationsfile > simulations
```

This `simulations` file requires that the system boundaries are stored in the file `stdexch.txt`.

Check the degradability of all metabolites

```
| make-fulldegradability-simulationsfile > simulations
```

This `simulations` file requires that the system boundaries are stored in the file `stdexch.txt`.

Check the network on leaks

The following `simulations` file makes two checks:

- Can any metabolite be produced without any inward transport while any outward transport is allowed?
- Can any metabolite be degraded without any outward transport while any inward transport is allowed?

This check is very similar to the check proposed by [5].

```
| make-leakcheck-simulationsfile > simulations
```

The function also writes two additional files, `any_outwards.txt` and `any_inwards.txt`, from all metabolites.

Exclude that metabolites can be created without substrates

```
| make-futileproducibility-simulationsfile > simulations
```

This `simulations` file requires that the excretable products stored in the file `stdwaste.txt`.

This is a weaker test as the leak test in the previous section since only some given waste products are allowed as excretion products and not any metabolite. It might still be a worthwhile check if the results of the former are difficult to interpret.

Exclude that metabolites can be degraded without excretion products

```
| make-wastability-simulationsfile > simulations
```

This `simulations` file requires that all possible substrates are stored in the file `stdsubstr.txt`.

5.6 Functions creating common exchange files referenced in a simulations file

Access the system boundaries in the original FABASE model

```
| make-standard-exchangables > stdexch.txt
```

This functions writes the metabolite marked as “open boundary” in the original FABASE model with the qualifier “=”, meaning it can be exchanged in both directions. If this model was created with `sbml2fa` it is exactly the boundary condition of the original SBML file.

```
| make-standard-wastables > stdwaste.txt
```

does the same but adds the qualifier “+” meaning that these substances can be excreted to be used by a `simulations` file created by `make-futileproducibility-simulationsfile`.

```
| make-standard-substrates > stdsubstr.txt
```

does the same but adds the qualifier “-” meaning that these substances can be imported to be used by a `simulations` file created by `make-wastability-simulationsfile`.

5.7 Network printing functions

The present FASIMU or FASBASE network can be converted in other formats. Some of these functions have already been executed and their results are stored in the current directory:

File	format	generating function
<code>reactions.txt</code>	plain text format	<code>reactions-printout equilibriums</code>
<code>reactionsa.txt</code>	dito, stoichiometrix factor 1 omitted	<code>reactions-printout equilibriums condensed reanames</code>
<code>reaclear.txt</code>	text format, metabolite names instead of identifiers	<code>reactions-printout equilibriums cleartext</code>
<code>reacleara.txt</code>	dito, stoichiometrix factor 1 omitted	<code>reactions-printout equilibriums condensed cleartext reanames</code>
<code>network.sbml</code>	SBML version 2 level 4	<code>print-sbml</code>

All functions print their output to the screen. They can be used both in a FASIMU or FABASE session. They do not print the pseudo reactions which FASIMU adds.

Plain text format: `reactions-printout`

This functions writes plain text format of the recent network. It is a tab-separated file. The first column contains the reaction identifier, the second column the reaction equation, separated by spaces. It is written to standard output, to save it to a file, call

```
| reactions-printout > network.txt
```

More columns are possible added by the options as follows.

cleartext Write metabolite name instead of identifiers.

condensed Suppress the stoichiometric coefficient.

equilibriums Write the equilibrium constant in the second column.

reanames Add a last column with the respective content of **reaction-names**.

SBML format: `print-sbml`

This functions writes a SBML version 2, level 4 to the standard output, to save it to a file use:

```
| print-sbml > network.sbml
```

Options:

noconcentrations Normally the concentrations stored in **concentration-class-ranges** in conjunction with **concentration-ranges-classes** are stored in a `<listOfConstraints>` section. The soft bounds result in a warning if exceeded, whereas the hard bound cause an error message. The set points values are written as the initial concentration variable in the species tag. This options suppresses this altogether.

nospeciotype Normally species are assigned a `speciesType`, the metabolite identifier and name without the compartment identifier. This options suppresses this.

noequilibriums Normally the equilibrium constants are stored in a tag: `<parameter id="equilibrium_constant" value=".."/>` inside a `<listOfParameters>` in a `<kineticLaw>` section for every reaction. This options suppresses this.

METATOOL format: `print-metatool-network`

Simple function to transform the current network into METATOOL format, similar to the above functions. It has no parameters.

expa format: `print-expa-network`

Simple function to transform the current network into expa format, similar to the above functions. It has no parameters.

Chapter 6

Functions for sets of modes: **modeset**

This chapter describes functions which operate on a set of flux distributions (modes). They have a special data format called **modeset**:

- tab separated
- line with no tab sign is the identifier of a mode
- line with a tab sign assigns a flux rate (second column) to a reaction identifier (first column)
- must start with a line of a mode identifier
- zero fluxes may be omitted (and should be)

Functions operating with this data type contain the word **modeset**.

6.1 Creating a **modeset** from **allout.txt** and vice versa

Normally you don't have to create this **modeset** by hand, the function **allout2modeset** creates one from the flux solutions in a FASIMU simulations run. The identifier of the mode is the name of the simulation. Beware of simulation runs where the same simulation identifier appears multiple times. There are different ways to call this function:

```
| allout2modeset
```

uses the file **allout.txt** and creates the file **modeset.txt**. You can also set the file names manually, in the following ways — each call has the same effect:

```
| allout2modeset allout.txt -o modeset.txt
| allout2modeset allout.txt -f > modeset.txt
| allout2modeset
| allout2modeset allout.txt
| allout2modeset -o modeset.txt
| allout2modeset -f > modeset.txt
```

The opposite direction is also possible, to create an **allout.txt** from a **modeset**. This is useful if you want to visualize the modes using the function **allout2bina**. Some information of the **allout.txt** coming from a simulation run is obviously not included: the description of the optimization call, the affirmation of the solver success, possible warnings of the solver. But for the functions that use **allout** that does not matter. Again there are different ways to call the function, the default file names are again **modeset.txt** and **allout.txt**.

```
| modeset2allout
```

uses the file `allout.txt` and creates the file `modeset.txt`. You can also set the file names manually, in the following ways — each call has the same effect:

```
| modeset2allout modeset.txt -o allout.txt
| modeset2allout modeset.txt -f > allout.txt
| modeset2allout -o allout.txt
| modeset2allout -f > allout.txt
| modeset2allout modeset.txt
| modeset2allout
```

6.2 Which mode fits a given profile best? `modeset-score`

This function returns a score (from 0 to 1) for each mode M_k in a given modeset how well it matches a given profile $V = (v_i)_i$. The profile V can be a reference flux distribution but also an expression pattern of protein or RNA abundance. The key for its biological usefulness is the correct combination of parameters. While some parameters must be set according to the type of reference profile, others have to be adjusted based on heuristics or trial and error. The parameters are described in the sequel.

6.2.1 Zeros in the mode M_k **sigscore** (-S) vs. **fullscore**

By default, the scoring includes all reactions. The total score for the pair M_k and V is by the arithmetic mean of single scores $\text{score}_i(m_i, v_i)$:

$$\text{Score}(M_k, V) = \frac{1}{n} \sum_{i=1}^n \text{score}_i(m_i, v_i)$$

where n is the number of reactions in the systems. This is called **fullscore**. However, **modeset-score** can also be configured to ignore the $m_i = 0$, where m_i are the components of $M_k = (m_i)_i$. It is called **sigscore**, the respective option is **-s**. It changes the score to

$$\text{Score}(M_k, V) = \frac{1}{\#I_n} \sum_{i \in I_n} \text{score}_i(m_i, v_i), \quad \text{where } I_n = \{i \mid m_i \neq 0\},$$

in other words, zero flux rates are ignored in this case.

The **sigscore** setting should be applied if the modes in the modeset are sparse, e.g. elementary flux modes [19] or minimal flux modes [6]. If the modes in the modeset are reference flux distributions representing the full metabolism at a given time point, **fullscore** should be preferred.

6.2.2 Scaling factor: fixed (-f), dependent on m_z (-L), or computed by linear regression

The score of M_k with respect to V is not applied directly but with a scaling factor λ . This is particularly relevant for the fitting setting (see below). So in fact, M_k is compared to λV . This factor can be set directly with **-f** in which case it is equal for all modes in the modeset. It is up to the user to ensure that this makes sense.

It can also be set with the formula

$$\lambda = l m_z$$

where l is a positive number set by the option **-L** (default is 1) and the formula for m_z (the average absolute value of the m_i) is given below.

Otherwise it is automatically computed with the formula:

$$\lambda = \frac{\sum_{i=1}^n v_i m_i}{\sum_{i=1}^n v_i^2} \quad \text{and for sigscore} \quad \lambda = \frac{\sum_{i \in I_n} v_i m_i}{\sum_{i \in I_n} v_i^2}$$

The summation is affected by the parameter **sigscore** in which case the i with $m_i = 0$ are ignored.

It is the explicit solution of the linear regression of M_k with V minimizing the EUKLIDIAN distance from M_k to λV . In case λ is zero, $\text{Score}(M_k, V)$ is set to zero. For the **absolute** setting (see below) and $\lambda < 0$, $\text{Score}(M_k, V)$ is also set to zero.

6.2.3 Relative (-R) vs. absolute

If the probe profile V is derived from the difference of two experiments it has to be considered relative (choose option **-r** in this case), otherwise it is considered absolute. This setting also affects how the modeset (M_k) is interpreted. In the relative case a reference mode of the modeset can be considered with a negative coefficient λ . In this case its interpretation is: “less of this mode”. In the absolute case this is not allowed.

Furthermore, in the **absolute** setting the values are considered by their absolute values. For the sake of brevity, only m_i will be written, but in the **absolute** setting $|m_i|$ is used. Think of it as the assignment:

$$M_k := |M_k|$$

6.2.4 Cumulative (-C) vs. fitting

For **cumulative** setting and for nonzero m_i the higher the expression value, the higher the score. Otherwise in the **fitting** setting it is maximal (and has value 1) at the reference point m_i/λ and decreases for both lower and higher values. The score distribution for the **cumulative** setting has its turning point and the 0.5 value at m_i/λ if it is not set by the **-c** option.

As a general rule, the **cumulative** setting is applicable to absolute protein or RNA expression profiles, while the **fitting** will usually be applied if the probe is a reference flux distribution (measured or inferred) or for relative probes. But there may be exceptions to this rule. The critical question is whether the score should monotonously increase for high expression values.

6.2.5 Setting the center of the score distribution manually (-c)

Normally, the center of the score distribution is m_i/λ set up for each M_k and each m_i individually. With this option you can set it up to a predefined value.

This apparently makes only sense in the **cumulative** and **absolute** setting for expression profiles which are normalized and the best splitting value for “on” and “off” genes is already known. This threshold value can be entered here.

6.2.6 Setting the steepness of the distribution function (-s)

This parameter sets the steepness of the distribution function, the higher the value the smaller the score variance. The smaller the variance the smaller the range of the significant score amplitude changes. The exact mathematical definition depends on the **type** parameter, for the binary **type** it is not used. A setting of one gives a good default steepness. The steepness value will be denoted s below.

6.2.7 Should the score variance depend on the flux value (varsingle -v)?

In the default case the score variance does not depend on the absolute value of $|m_i|$, it is equal for each reaction and is computed from the average absolute value of M_k :

$$m_z = \frac{1}{n} \sum_{i=1}^n |m_i| \quad \text{and for sigscore} \quad m_z = \frac{1}{\#I_n} \sum_{i \in I_n} |m_i|.$$

With the setting `-v` the variance of nonzero m_i depends linearly from $|m_i|$ instead of m_z . Only for $m_i = 0$ it is m_z .

This setting `-v` is recommended if the absolute values of the flux rates in M_k is very unevenly distributed. If an average variance is then used, the values with smaller absolute values will have a score distribution which goes in the opposite direction.

The exact mathematical definition depends on the `type` parameter, for the binary `type` it is not used.

6.2.8 Mathematical description of the score distribution (-t)

Possible values are (where `gauss` is the default):

`-t gauss`

The score is in the fitting case the GAUSSIAN bell curve of the normal distribution with the mean m_i/λ and the standard deviation of sm_z/λ scaled to equal 1 at the maximum, where s is the steepness parameter:

$$\text{score}_i(m_i, v_i) = e^{-\frac{1}{2} \left(s \frac{\lambda v_i - m_i}{m_z} \right)^2}$$

except for the `varsingle` setting and $m_i \neq 0$ where it is:

$$\text{score}_i(m_i, v_i) = e^{-\frac{1}{2} \left(s \frac{\lambda v_i - m_i}{|m_i|} \right)^2}$$

In the cumulative it is for $m_i \neq 0$

$$\text{score}_i(m_i, v_i) = \Phi \left(s \frac{\lambda v_i - m_i}{m_z} \right)$$

and for $m_i = 0$

$$\text{score}_i(m_i, v_i) = 1 - \Phi \left(s \frac{\lambda v_i - m_i}{m_z} \right)$$

For the `varsingle` setting and $m_i \neq 0$ it is:

$$\text{score}_i(m_i, v_i) = \Phi \left(s \frac{\lambda v_i - m_i}{|m_i|} \right)$$

`-t cosine`

The score is in the fitting case a half period of the cosine function

$$\text{score}_i(m_i, v_i) = \frac{1}{2} \left(1 + \cos \left(s \frac{\lambda v_i - m_i}{m_z} \frac{\pi}{2} \right) \right)$$

if v_i is in the range $(\frac{m_i}{\lambda} \pm \frac{2m_z}{s\lambda})$ and zero otherwise.

For the `varsingle` setting and $m_i \neq 0$ it is:

$$\text{score}_i(m_i, v_i) = \frac{1}{2} \left(1 + \cos \left(s \frac{\lambda v_i - m_i}{|m_i|} \frac{\pi}{2} \right) \right)$$

if v_i is in the range $(\frac{m_i}{\lambda} \pm \frac{2|m_i|}{s\lambda})$ and zero otherwise.

In the cumulative case it is for $m_i \neq 0$

$$\text{score}_i(m_i, v_i) = \frac{1}{2} \left(1 + \sin \left(s \frac{\lambda v_i - m_i}{m_z} \frac{\pi}{4} \right) \right)$$

if v_i is in the range $(\frac{m_i}{\lambda} \pm \frac{2m_z}{s\lambda})$, 1 if it is above, and 0 if it is below the range, and for $m_i = 0$ it is

$$\text{score}_i(m_i, v_i) = \frac{1}{2} \left(1 - \sin \left(s \frac{\lambda v_i - m_i}{m_z} \frac{\pi}{4} \right) \right)$$

if v_i is in the range $(\frac{m_i}{\lambda} \pm \frac{2m_z}{s\lambda})$, 0 if it is above, and 1 if it is below the range.

For the varsingle setting and $m_i \neq 0$ it is:

$$\text{score}_i(m_i, v_i) = \frac{1}{2} \left(1 + \sin \left(s \frac{\lambda v_i - m_i}{|m_i|} \frac{\pi}{4} \right) \right)$$

if v_i is in the range $(\frac{m_i}{\lambda} \pm \frac{2|m_i|}{s\lambda})$, 1 if it is above, and 0 if it is below the range.

-t parabel

The score is in the fitting case the top of an inverted parabel:

$$\text{score}_i(m_i, v_i) = 1 - \min \left\{ 1, \left(s \frac{\lambda v_i - m_i}{2m_z} \right)^2 \right\}$$

For the varsingle setting and $m_i \neq 0$ it is:

$$\text{score}_i(m_i, v_i) = 1 - \min \left\{ 1, \left(s \frac{\lambda v_i - m_i}{2|m_i|} \right)^2 \right\}$$

In the cumulative case it is for $m_i \neq 0$

$$\text{score}_i(m_i, v_i) = 1 - \max \left\{ 0, \min \left\{ 1, \frac{1}{2} \left(s \frac{\lambda v_i - m_i}{2m_z} \right)^3 \right\} \right\}$$

and for $m_i = 0$ it is

$$\text{score}_i(m_i, v_i) = \max \left\{ 0, \min \left\{ 1, \frac{1}{2} \left(s \frac{\lambda v_i - m_i}{2m_z} \right)^3 \right\} \right\}$$

For the varsingle setting and $m_i \neq 0$ it is:

$$\text{score}_i(m_i, v_i) = 1 - \max \left\{ 0, \min \left\{ 1, \frac{1}{2} \left(s \frac{\lambda v_i - m_i}{2|m_i|} \right)^3 \right\} \right\}$$

-t binary

The binary function is much simpler, the steepness parameter s has no effect, λ is neither computed or set.

The score is in the fitting case is

$$\text{score}_i(m_i, v_i) = \frac{1}{2} (\text{sgn} v_i + \text{sgn} m_i)$$

and in the cumulative case t is

$$\text{score}_i(m_i, v_i) = \min \left\{ 1, \frac{1}{2} (2 + \text{sgn} v_i - \text{sgn} m_i) \right\}.$$

In the relative case $\text{score}_i(m_i, -v_i)$ is also computed and used if is larger than $\text{score}_i(m_i, v_i)$.

Exclude reactions from scoring: -x

This option removes the respective reactions from the computation of $\text{Score}(M_k, V)$ and also the computation of λ and m_z . The option must be called with a filename containing one reaction identifier per line. The option can appear multiple time with different file names: in this case the reaction identifier of all files are regarded. The computation of $\text{Score}(M_k, V)$ with the set X of excluded indeces changes to:

$$\text{Score}(M_k, V) = \frac{\sum_{i \notin X} w_i \text{score}_i(m_i, v_i)}{\sum_{i \notin X} w_i} \quad \text{and for sigscore} \quad \text{Score}(M_k, V) = \frac{\sum_{i \in I_n \setminus X} w_i \text{score}_i(m_i, v_i)}{\sum_{i \in I_n \setminus X} w_i}$$

Assigning a weight to reactions: -w

By default, every reaction carries the same weight in the computation of $\text{Score}(M_k, V)$, λ and m_z . This option reads a file with weights in the following format:

- whitespace separated
- first token: reaction identifier of reaction i
- second token: non-negative number w_i .

The computation of $\text{Score}(M_k, V)$ changes to:

$$\text{Score}(M_k, V) = \frac{\sum_{i=1}^n w_i \text{score}_i(m_i, v_i)}{\sum_{i=1}^n w_i} \quad \text{and for sigscore} \quad \text{Score}(M_k, V) = \frac{\sum_{i \in I_n} w_i \text{score}_i(m_i, v_i)}{\sum_{i \in I_n} w_i}$$

Reaction identifiers not appearing in the weight files are assumed to have weight 1. A weight of zero is also allowed, it has the same effect as including the respective reaction in the excluded file (-x).

Assigning a flux dependent weight to reactions: -W

This option assigns another (multiplicative) weight component which depends on the flux value m_i . The option allows an optional parameter τ (default 1).

The computation of $\text{Score}(M_k, V)$ changes to:

$$\text{Score}(M_k, V) = \frac{\sum_{i=1}^n |m_i|^{1/\tau} \text{score}_i(m_i, v_i)}{\sum_{i=1}^n |m_i|^{1/\tau}} \quad \text{and for sigscore} \quad \text{Score}(M_k, V) = \frac{\sum_{i \in I_n} |m_i|^{1/\tau} \text{score}_i(m_i, v_i)}{\sum_{i \in I_n} |m_i|^{1/\tau}}$$

If both weight options -w and -W are given the weight components are simply multiplied:

$$\text{Score}(M_k, V) = \frac{\sum_{i=1}^n w_i |m_i|^{1/\tau} \text{score}_i(m_i, v_i)}{\sum_{i=1}^n w_i |m_i|^{1/\tau}} \quad \text{and for sigscore} \quad \text{Score}(M_k, V) = \frac{\sum_{i \in I_n} w_i |m_i|^{1/\tau} \text{score}_i(m_i, v_i)}{\sum_{i \in I_n} w_i |m_i|^{1/\tau}}$$

Ignore the reference flux: -i

This option ignores the flux value in a way that m_i and m_z is set to 1 in the score formula but not in the computation of the weight in the -W option.

Displaying the scores for single reactions: -D

The output (showing the total score) is preceded by a tab-separated table showing

- the reaction identifier,
- the expression value,
- the scaled expression value,
- the flux value in the reference flux mode,
- the computed score,
- the weighted computed score, and
- the weight.

There is also the option `-d` showing additional (debug) information which is not documented in detail here.

Chapter 7

Using FABASE without FASIMU

The question arises why the two layer structure FABASE vs. FASIMU which resulted from historical reasons is still retained. The answer is that it may still be worthwhile to use FABASE on its own so this possibility is still supported.

To use FABASE without FASIMU is a reasonable alternative if FASIMU's simulations concept is not needed, i.e. if the focus is on a single optimization problem (biomass maximization for instance). Reasons might also be related to computation times: FASIMU increases the number of reactions and metabolites considerably (one extra reaction and metabolite for each metabolite) to control the systems boundaries. The intrinsic problem complexity is not changed by this modification, however, the files get larger, and some scripts may be affected by this.

The main reason however to use FABASE alone is for experimentation. In a typical FASIMU session, the basic information files (the LPF-files) are overwritten frequently and some flow of information is not visible. It may even be sensible to modify certain LPF-files in a text editor to obtained algorithmic features not yet (or never be) implemented in FASIMU. Also a combined approach is possible: start with FASIMU (including the added system boundary control) and continue with FABASE functions.

Technically, the main difference between a FASIMU and FABASE session is that in a FABASE session you call `compute-FBA` (or other `compute-` functions) directly from the command line where in a FASIMU session you put the respective `compute-` functions into the variable `$optimization_call` and enter `simulate` or `simulate_single` in the command line — the `compute-` functions are called indirectly. The other important difference is that in a FABASE session you have to put the objectives in the file `targetfluxes` and the constraints in the file `fluxconstraints` where in a FASIMU session the objectives and constraints are condensed in a single line of `simulations` and `targetfluxes` and `fluxconstraints` are written upon this information. Simply put: FABASE: more control but also more manual work. The startup of a FABASE session is:

```
| source fasbase
```

One example of a small FABASE session is given in the tutorial about *E. coli*: the modification of the equilibrium constants into a set obeying the WEGSCHEIDER condition. Here, the function `well-formed-equilibriums` is called directly on the model.

7.1 Computation functions

The principal optimization function: `compute-FBA`

The more comprehensive description can be found in section 4.3. Here, a more technical description can be found. The basic usage is:

| compute-FBA [options]

compute-FBA on its own is translated to compute-FBA -F. Description of the options follow.

Options for flux minimization: -F [<fluxmin-type>]

Type	weight of forward flux	weight of backward flux
h	1	K_{eq}
0	1	1
1	$\frac{1}{1+K_{eq}}$	$\frac{K_{eq}}{1+K_{eq}}$
2	$\frac{1}{1+K_{eq}^2}$	$\frac{K_{eq}}{1+K_{eq}^2}$
s	$\frac{1}{\sqrt{1+K_{eq}^2}}$	$\frac{K_{eq}}{\sqrt{1+K_{eq}^2}}$
e	<enzyme cost>	<enzyme cost>
E	<enzyme cost>	<enzyme cost> K_{eq}
m	1 if $v = 0$, 0 otherwise	

The type can be omitted in which case it is 0 if -T is also set, and h otherwise.

Specific minimization: --m [<rea-ID>] ...

The flux of the given reaction(s) is minimized. For this to be useful it is necessary that at least one flux is set to a nonzero value (e.g. with the **setfluxes** file), otherwise the zero solution will be computed.

Specific maximization: --b [<rea-ID>] ...

The flux of the given reaction(s) is maximized. For this to be useful it is necessary that wither the intake or the output fluxes are restricted (e.g. with the **fluxbounds** file), otherwise the problem becomes unbounded.

Fitness maximization: --f [<type> [<weight>]]

Type	Norm	Adjustment	Fitness score	Restriction
0	Euklidian	none	$1 - \sqrt{\frac{\sum \Delta v_i^2}{\sum L_i^2}}$	$\text{sgn}(v_i) = \text{sgn}(L_i)$
1	Euklidian	partly (linear)	$1 - \sqrt{\frac{\sum \frac{\Delta v_i^2}{ L_i }}{\sum L_i }}$	
2	Euklidian	full	$1 - \sqrt{\frac{1}{n} \sum \frac{\Delta v_i^2}{L_i^2}}$	
3	linear	full	$1 - \frac{1}{n} \sum \frac{ \Delta v_i }{ L_i }$	
4	linear	none	$1 - \frac{\sum \Delta v_i }{\sum L_i }$	
5	linear	full	$1 - \frac{1}{n} \sum \frac{ \Delta v_i }{ L_i }$	
6	maximum	none	$1 - \frac{\max\{ \Delta v_i \}}{\max\{ L_i \}}$	
7	maximum	full	$1 - \max\left\{\frac{ \Delta v_i }{ L_i }\right\}$	
8	quadratic	full	$1 - \frac{1}{n} \sum \frac{\Delta v_i^2}{L_i^2}$	

The **weight** is multiplied with the above coefficient. Fitness maximization can be combined with flux minimization and specific minimization but not with maximization.

Using expression profiles

This function requires the file **expressions** to be present, giving the expression values. The call is:

| compute-FBA -x [<weight>] [-xu <upper>] [-xl <lower>] [-xs <significant>]

The option `-x` switches the function on, meaning that specific terms are added to the scoring function. Thus, this feature can be combined with other optimizations. Defaults are

Parameter	meaning	default value
<weight>	relative weight with respect to scoring function	1000
<upper>	upper threshold on positive expression	0.5
<lower>	lower threshold on negative expression	0.49
<significant>	threshold on the significance of the absolute value of a flux	1

Thermodynamic realizability

Options for TR:

Option	purpose	default value
-T	use TR criterion, default type	A 1 10^{-5}
-T [[A-Ea-e]r?]	use TR with type identifier	
-s [<weight>]	use soft bounds	
-w [<weight>]	use set points	
-k <number>	potential tolerance maximum	
-K <number>	penalty multiplicator for potential tolerance	
<num>..<>	alternative way to enter concentration values	

TR types are summarized in this table:

Type	Zero flux	Zero potential	Irreversible reaction	comment
A	q arbitrary	any flux	potential sign fixed	Conditional clause implementation
B	$q = 0$	$v = 0$		Conditional clause implementation
a	q arbitrary	any flux		bigM implementation
b	$-\epsilon < q < \epsilon$	$-\epsilon < v < \epsilon$		bigM implementation
Ar	q arbitrary	any flux		Conditional clause implementation
Br	$q = 0$	$v = 0$		Conditional clause implementation
ar	q arbitrary	any flux		bigM implementation
br	$-\epsilon < q < \epsilon$	$-\epsilon < v < \epsilon$		bigM implementation

Program control options

Option	purpose
-p	prepare only 'problem.lp', do not compute
-t <seconds>	timeout in seconds, finish (single flux computation), default 300, 0 means unlimited time
-c	check the results with another cplex run
-d	print documentation files for thermodynamic feasibility
-CPLEXnodes <nodes>	abolish computation number of nodes, default 5000, 0 means unlimited number of nodes (CPLEX only)
-CPLEXemph <1..4>	MIP emphasis setting to be tried out in critical size models (CPLEX only)
-CPLEXline "<param-line>"	Give this line to CPLEX

7.2 Further functions in FABASE

Ensure standard directions

The function `ensure-standard-directions` ensures the convention that the reactions are written such that the standard Gibb's free energy is negative, i.e. the equilibrium constant is larger or equal to 1. In other words, if all metabolites have a concentration of 1M each reaction proceeds in forward direction. This condition is assumed in the formulation of the flux minimization algorithm [8], which is implemented as

`compute-FBA -F h` in FASIMU. The function simply exchanges products and substrates in the reaction equation for the respective reactions, also modifies the boundaries stored in the file `reactions` accordingly, and replaces the equilibrium constant by its inverse. It does currently not change the contents of `setfluxes`, `fluxconstraints`, `fluxbounds`, `fluxfix` etc. . The user has to take of that.

The function modifies the files `reactions` and `equilibriums`. However, it does not regenerate all information depending on these files. Therefore, it does not work just to continue with the FASIMU/FABASE session. The recommended way to use the function is to create a new model with the modified files `reactions` and `equilibriums`;, as follows, assuming the original model is in `$modelldir/model1`:

```
mkdir newdir
cd newdir
cp -R $modelldir/model1/*
source fabase
ensure-standard-directions
cp -R $modelldir/model1 $modelldir/model2
cp reactions equilibriums $modelldir/model2
rm *
cd ..
rmdir newdir
```

Still, it is possible to restart a FABASE session with modified `reactions` file, just call `fabase-main`. Generally, it is not a good idea to modify the file `reactions` in a FASIMU session, since `fasimu` modifies the model, but it is possible with:

```
ensure-standard-directions
rm reactions.original equilibriums.original
fasimu-main
```

Well-formed equilibriums

For thermodynamic computations it is favorable if the standard reaction free energies are compatible to each other, in other words, they obey the WEGSCHEIDER condition. Experimental values from different sources obviously do not exactly comply with this rule. Values derived from formation energies by prediction methods [13] should do so but this is not always the case due to computational errors. The function `well-formed-equilibriums` attempts to force this condition with changes to the data given in `equilibriums`. It is done with an error minimization which is either linear (`-l` option) or quadratic (`-q` only available for solver CPLEX). The latter usually spreads the modification values on more reactions. This algorithm is described in [11]. There are two files modifying the algorithm: If a reaction is included in the file `trusted-equilibriums` it will not be modified. If a reaction is included in the file `untrusted-equilibriums` it will primarily be changed. The corrected set of equilibrium constants will be written to `corrected-equilibriums` and a modification report is written to `eq-corr-report.txt`. It is recommended not continue the FABASE or FASIMU session with a modified `equilibriums` file but to generate a separate model instead as described in the previous section (Ensure standard directions). However, in a FABASE session, `fabase-main` can be called, or in FASIMU session `fasimu-main` can be called to regenerate dependent information.

Another recommended way to modify the model prior to a FASIMU session is to start a small `fabase` session just for the modification of the model, and then start FASIMU in the normal way:

```
source fabase
well-formed-equilibriums
source fasimu
```

This also works with `restrict-equilibriums` and `ensure-standard-directions`.

Remove duplicates

During network reconstruction work, occasionally the same reaction is recorded twice. It might be easily overlooked if the order of reactants is different and substrates/products are reversed. Duplicated reactions can also be the result if two compartments are automatically merged.

Normally, duplicates do not present problems in the computation — just one of the reaction carries the flux. But it might lead to confusion in the interpretation of the result. Troublesome can be the situation when differing extra annotation is assigned to the reactions. Thus, it is advisable to check for duplicates frequently in the curation process.

The function `remove-duplicates` performs the following steps:

- for each reaction, the substrates and the products are ordered lexicographically (you can inspect the result in the text file `reactions_ref.fgf`).
- Each reaction is compared with each other reaction, directly and with products and substrates reversed. (In fact, a more efficient method is used using `gawks` hashlist array.) If it is found it is recorded in the file `reactions_redundant.fgf` (except for a situation described below).

That is a tab-separated file: the first two columns hold the respective reaction identifiers. The first identifier is the one whose reaction is prepared to be retained in the network the second one belongs to the reaction to be deleted. In a more than two-way equality also reactions belonging to the first identifier in a row might be deleted. The third column holds the keyword forward or backward (indicating if the product/substrate sides had to be reversed), and the fourth holds a comment (see the following point).

The flux bounds of both reactions are checked. If they do not overlap the reaction pair is not recorded. For instance if forward and backwards reactions are split they will not be removed by remove duplicates. If one flux interval is a subset of the other the more specified reaction is recorded in the first column (comment “by stricter bounds”). If the flux intervals overlap but are not subsets, the reactions with the higher value of the lower bound chosen for the first column (comment “by stricter lower bound”). If the flux intervals are equal the lexicographically first entry is chosen for the first column (comment “lexicographic for identical reactions”).

- The identifiers of the second column are collected into a single file: `removables.fgf`.

The function `remove-duplicates` does *not* remove the reactions right away for obvious reasons: it is worthwhile to check the reactions to be removed. The function `apply_removables` finally removes the reactions from the file `reactions` and restarts FABASE. It uses the file `removables.fgf` for that purpose. If the user want to delete other reactions, the file `removables.fgf` should be modified before calling `apply_removables`.

Adjust metabolites file according to reactions file

Normally, FASIMU requires that all metabolites referenced in `reactions` are defined in the file `metabolites` and each entry in `metabolites` appears as a reactant in `reactions`. If that is not the case from the start the file `metabolites` is automatically modified with the function `correct-metabolites`. Each missing metabolite is added to `metabolites` (the name is the identifier, and it is considered “closed boundary”), and extra metabolites are removed. For each modification, a warning is printed to the screen. You can also call this function manually to check and to see the warnings. However, the function is also called automatically by `source fasimu`, `source fabase`, `fabase-main` etc. .

Filter reactions

In the computation with large networks, often a subnetwork is sufficient to model certain metabolic functions. It is possible to duplicate the large network and remove the unnecessary reactions from the network. But

if errors become apparent in the large network and it is modified later the submodels are still contain these errors. To repair them, the modifications can either be applied to every submodel created or the duplication process can be repeated. Both methods are quite cumbersome.

FASIMU offers a more elegant way to deal with this kind of model reduction. Unnecessary parts of the network can usually easily be defined by set of reactions or metabolites.

The function `reactions-filterout` takes one argument, a file with a list of reaction identifiers, and removes them from the file `reactions`. If there are metabolites which are not occurring in any reaction they are removed automatically from the file `metabolites`. This is done by calling the function `metabolites-correct`. Finally, `fabase-main` is called which restarts the FABASE session.

The function `metabolites-filterout` also takes one argument, a file with a list of metabolite identifiers, and removes every reaction from the file `reactions` in which one of these metabolites occur. Consequently, these metabolites are also removed from the file `metabolites`.

These functions should not be called in a FASIMU session as they modify only `reactions` and `metabolites` but not `reactions.original` and `metabolites.original`. The purpose of the functions is to adjust the model in a FABASE session *before* the FASIMU session is started. The recommended way to reduce the model before computation is:

```
source fabase
reactions-filterout noreas.txt
metabolites-filterout nomets.txt
source fasimu
...
```

7.3 Modify model files in a FABASE session

The recommended way to handle your models

Generally, I do not recommend to modify the files in the directory in which you are working with FABASE. You should keep the original models in separate directories and call `source fabase` in a fresh directory. Say, you keep and modify your model in a directory `~/model` then proceed like that:

```
cd ~
cp -R model model-compute
cd model-compute
source fabase
<do the computations>
<copy the result files to a designated directory>
cd ..
rm -r model-compute
```

Hoever, for experimentation you can change all input files during the session. The program is capable to regenerate intermediate files but that requires to call functions.

Modification functions

See the following table for the modification functions. If you change one of items below the function on the right hand side must be called. `fabase-main` regenerates every intermediate file, so it can be used if several files are changed.

Modified file	possible	Function for the safe integration of the modification
reactions	yes	fabase-main
metabolites	yes	fabase-main
equilibriums	yes	fabase-main
TR-excluded	yes	update-TR-excludes
fluxmin-excluded	yes	update-enzymes (only if enzymes are used)
fluxmin-weights	yes	update-enzymes (only if enzymes are used)
concentration-ranges-classes	yes	update-concentration-ranges
concentration-class-ranges	yes	update-concentration-ranges
fluxbounds	yes	update-fluxbounds
setfluxes	yes	update-fluxbounds
fluxconstraints	yes	update-fluxbounds
targetfluxes	yes	make-vset-lpf
enzymes	yes	update-enzymes
fluxfix	yes	update-fluxfix
expressions	yes	not necessary
cplex-tail.in	yes	not necessary
lp_solve.par	yes	not necessary
lindo.par	yes	not necessary
\$glpk_opts	yes	not necessary

Chapter 8

Customization of the external solvers

Large networks in combination with computational expensive constraints such as

- Thermodynamic realizability (requires Boolean variables, thus, computing a flux distribution is now at least a mixed-boolean linear problem, and the number of variables increases: for each metabolite one Boolean and one real-valued variable)
- Set points for concentration values (`-w` option) turn the problem into a quadratic objective problem
- Use of the fitness maximization, types 0...2 also turns the problem into a quadratic constraint problem
- stoichiometric coefficients are real numbers or have a large value (increase the numerical difficulty of the problem, which might cause apparently wrong solutions)

may push the optimization on the brink of their capability. However, all solvers can be customized. Choosing the right parameter switch may put a previously unsolvable problem into reach. You should inspect the file “`solver.out`” which is the output of the solver program stored after each computation call by FASIMU and the instruction manual of the respective solver.

By default, FASIMU instructs the solver to finish a single optimization after 5 minutes. The functions

```
| set-timeout <seconds>
```

modifies the configuration of the active solver to change this time span. The value 0 disables the timeout feature.

CPLEX

As FASIMU has been developed with CPLEX, a few default settings have been found to ideally harmonize with difficult FBA optimizations. The parameters are stored in the file `cpex-tail.in` which you can modify to change the values of parameters. You can display them with

```
| cat cpex-tail.in
```

The file will not be overwritten by a call of “`source fasimu`”; the changes will be in effect for all further computations in this directory. The defaults can be restored with:

```
| restore-defaults-cplex
```

LINDO

Lindo parameters are stored in “lindo.par”. You can display them with

```
| cat lindo.par
```

Again, this file will not be overwritten by a subsequent call of “source fasimu”; the defaults can be restored with:

```
| restore-defaults-lindo
```

lp_solve

Parameters for lp_solve are stored in lp_solve.par. You can display them with

```
| cat lp_solve.par
```

For more information on parameters see <http://lpsolve.sourceforge.net/5.5>. The file lp_solve.par will not be overwritten by a subsequent call of “source fasimu”; the defaults can be restored with:

```
| restore-defaults-lp_solve
```

GLPK

GLPK parameters are stored in the bash variable “\$glpk_opts”. The variable contains command line options given to GLPK. Call

```
| glpsol -h
```

to see a list of options. To apply different parameters you have to adjust this variable. You can display them with

```
| echo $glpk_opts
```

To see a list of possible parameters, call

```
| glpsol --help
```

in the command line. The variable is not changed by a subsequent call of “source fasimu”; the default can be restored with:

```
| restore-defaults-glpk
```

Chapter 9

Final remarks

9.1 Publication

If you found FASIMU useful for your scientific work you are asked to cite the publication:

Andreas Hoppe, Sabrina Hoffmann, Andreas Gerasch, Christoph Gille and Hermann-Georg Holzhütter. FASIMU: flexible software for flux-balance computation series in large metabolic networks. BMC Bioinformatics 2011, **12**:28. doi:10.1186/1471-2105-12-28, PMID:21255455.

9.2 Getting more help

There is also a FASIMU tutorial which may be helpful to you.

Functions help page

Some functions have their own help pages available with the “-h” option: `compute-FBA`, `compute-shlomi`, `well-formed-equilibriums`, `compute-moma`, `compute-room`. FASIMU has an online help page: `fasimu-help`.

Source code

FASIMU is written in the high-level languages bash and gawk so if you are fairly familiar with the syntax of these languages it may be worthwhile to look in the source code. For some functions additional information is available in the comments of the source code. The functions related to FABASE are defined in `/usr/local/bin/fabase`, related to FASIMU in `/usr/local/bin/fasimu` (for the recommended installation places).

FASIMU project website

You might wish inspect the project’s website at

<http://www.bioinformatics.org/fasimu>

for the latest updates and other information. It is highly recommended that you subscribe to the news list at

<http://www.bioinformatics.org/mailman/listinfo/fasimu-news>

to get informed on updates. Please report bugs to the bug tracking

http://www.bioinformatics.org/bugs/?group_id=1004

There is also a public forum at

http://www.bioinformatics.org/forum/?group_id=1004

9.3 License

FASIMU is published under the GNU public license (GPL) which can be viewed in a FASIMU session with

```
| fasimu-license
```

There is also a license statement at the beginning of each of the source files. This document is likewise under the GPL.

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